

PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

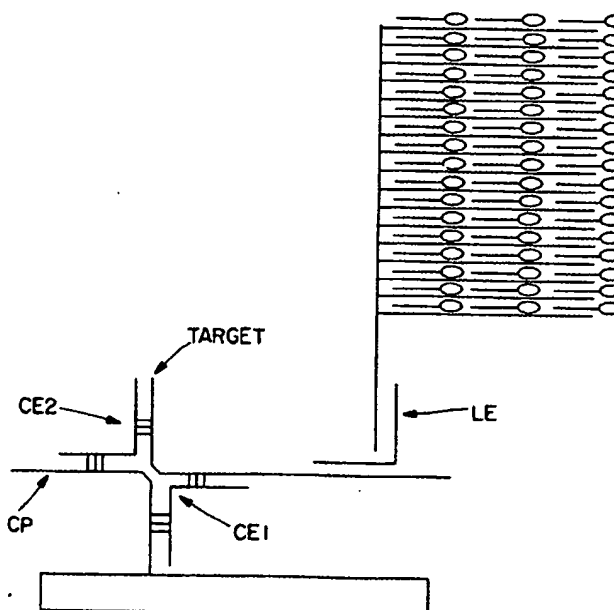
INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C12Q 1/68, C07H 21/00	A1	(11) International Publication Number: WO 95/16055 (43) International Publication Date: 15 June 1995 (15.06.95)
(21) International Application Number: PCT/US94/14119 (22) International Filing Date: 7 December 1994 (07.12.94) (30) Priority Data: 08/164,388 8 December 1993 (08.12.93) US (71) Applicant: CHIRON CORPORATION [US/US]; 4560 Horton Street, Emeryville, CA 94608 (US). (72) Inventors: URDEA, Michael, S.; 100 Bunce Meadow Road, Alamo, CA 94507 (US). FULTZ, Timothy; 358 Willow Creek Lane, Martinez, CA 94553 (US). WARNER, Brian, D.; 1034 Alhambra Avenue, Martinez, CA 94553 (US). COLLINS, Mark; 2991 Santos Lane #301, Walnut Creek, CA 94596 (US). (74) Agents: GOLDMAN, Kenneth, M. et al.; Chiron Corporation, Intellectual Property - R440, P.O. Box 8097, Emeryville, CA 94662-8097 (US).	(81) Designated States: AU, CA, HU, JP, KR, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>	

(54) Title: SOLUTION PHASE NUCLEIC ACID SANDWICH ASSAYS HAVING REDUCED BACKGROUND NOISE

(57) Abstract

New techniques are provided for substantially reducing background signals encountered in solution phase hybridization assays. The techniques are premised on eliminating or significantly reducing the phenomena of nonspecific hybridization and nonspecific binding, so as to provide a detectable signal which is produced only in the presence of the target polynucleotide of interest. In certain embodiments, methods are provided for increasing the signal which can otherwise be diminished in noise reduction. Kits for carrying out the novel assays are provided as well.



FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	GB	United Kingdom	MR	Mauritania
AU	Australia	GE	Georgia	MW	Malawi
BB	Barbados	GN	Guinea	NE	Niger
BE	Belgium	GR	Greece	NL	Netherlands
BF	Burkina Faso	HU	Hungary	NO	Norway
BG	Bulgaria	IE	Ireland	NZ	New Zealand
BJ	Benin	IT	Italy	PL	Poland
BR	Brazil	JP	Japan	PT	Portugal
BY	Belarus	KE	Kenya	RO	Romania
CA	Canada	KG	Kyrgyzstan	RU	Russian Federation
CF	Central African Republic	KP	Democratic People's Republic of Korea	SD	Sudan
CG	Congo	KR	Republic of Korea	SE	Sweden
CH	Switzerland	KZ	Kazakhstan	SI	Slovenia
CI	Côte d'Ivoire	LI	Liechtenstein	SK	Slovakia
CM	Cameroon	LK	Sri Lanka	SN	Senegal
CN	China	LU	Luxembourg	TD	Chad
CS	Czechoslovakia	LV	Latvia	TG	Togo
CZ	Czech Republic	MC	Monaco	TJ	Tajikistan
DE	Germany	MD	Republic of Moldova	TT	Trinidad and Tobago
DK	Denmark	MG	Madagascar	UA	Ukraine
ES	Spain	ML	Mali	US	United States of America
FI	Finland	MN	Mongolia	UZ	Uzbekistan
FR	France			VN	Viet Nam
GA	Gabon				

SOLUTION PHASE NUCLEIC ACID SANDWICH ASSAYS
HAVING REDUCED BACKGROUND NOISE

Technical Field

5 This invention relates generally to nucleic acid chemistry and hybridization assays. More particularly, the invention relates to methods for generating a more target-dependent signal in solution phase sandwich hybridization assays by minimizing background noise deriving primarily from
10 nonspecific hybridization and/or nonspecific binding. The invention additionally relates to methods for compensating for lost signal while reducing background noise. The invention further relates to kits containing the reagents necessary for carrying out the disclosed assays.

15

Background

 Nucleic acid hybridization assays are commonly used in genetic research, biomedical research and clinical diagnostics. In a basic nucleic acid hybridization assay,
20 single-stranded analyte nucleic acid is hybridized to a labeled single-stranded nucleic acid probe and resulting labeled duplexes are detected. Variations of this basic scheme have been developed to enhance accuracy, facilitate the separation of the duplexes to be detected from extraneous
25 materials, and/or amplify the signal that is detected.

 The present invention is directed to a method of reducing background signals encountered in solution phase sandwich hybridization assays that derive from several sources. Generally, the background noise which is addressed
30 by way of the presently disclosed techniques results from undesirable interaction of various polynucleotide components that are used in a given assay, i.e., interaction which gives rise to a signal which does not correspond to the presence or

- 2 -

quantity of analyte. The invention is useful in conjunction with any number of assay formats wherein multiple hybridization steps are carried out to produce a detectable signal which correlates with the presence or quantity of a polynucleotide analyte.

One such assay is described in detail in commonly assigned U.S. Patent No. 4,868,105 to Urdea et al. That assay involves the use of a two-part capturing system designed to bind the polynucleotide analyte to a solid support, and a two-part labeling system designed to bind a detectable label to the polynucleotide analyte to be detected or quantitated. The two-part capture system involves the use of capture probes bound to a solid support and capture extender molecules which hybridize both to a segment of the capture probes and to a segment of the polynucleotide analyte. The two-part labelling system involves the use of label probe extender molecules which hybridize to a segment of the polynucleotide analyte, and label probes which hybridize to the label probe extender molecules and contain or bind to a detectable label. An advantage of such a system is that a plurality of hybridization steps must occur in order for label to be detected in a manner that correlates with the presence of the analyte, insofar as two distinct hybridization reactions must occur for analyte "capture," and, similarly, two distinct hybridization reactions must occur for analyte labelling. However, there remain a number of ways in which a detectable signal can be generated in a manner which does not correspond to the presence or quantity of analyte, and these will be discussed in detail below.

Another example of an assay with which the present invention is useful is a signal amplification method which is described in commonly assigned U.S. Patent No. 5,124,246 to Urdea et al. In that method, the signal is amplified through the use of amplification multimers, polynucleotides which are constructed so as to contain a first segment that hybridizes specifically to the analyte nucleic acid or a strand of nucleic acid bound to the

analyte, and a multiplicity of second segments that hybridize specifically to a labeled probe. The degree of amplification is theoretically proportional to the number of iterations of the second segment. The multimers may be either linear or
5 branched. Branched multimers may be in the shape of a fork or a comb, with comb-type multimers preferred.

One approach which has been proposed to increase the target dependence of the signal in a hybridization assay is described in European Patent Publication No. 70,685,
10 inventors M.J. Heller et al. That reference describes a homogeneous hybridization assay in which a nonradiative transfer of energy occurs between proximal probes; two distinct events must occur for a target-generated signal to be produced, enhancing the accuracy of detection. A second
15 approach designed to enhance the signal deriving from the presence of analyte is described in European Patent Publication No. 361,983 inventor J.E. Stefano. The method described therein involves hybridization of two probe sequences (one a midvariant RNA (MDV), the other a half-
20 ribozyme), each of which is complementary to sequences present in an RNA target. The ribozyme thus formed specifically cleaves the tail of the MDV probe, releasing the MDV probe from the support and enhancing its replication. Still a third approach to increase specificity in hybridi-
25 zation assays is described by Distefano et al., J. Am Chem. Soc. (1992) 114:11006-11007. That method involves the use of short regions of double-helix formation to enhance the stability of two short regions of triple-helix DNA.

European Patent Publication No. 552,931,
30 inventors Hogan et al., describe a nucleic acid hybridization assay utilizing a probe system that detects regions of double-stranded DNA that only form in the presence of target sequence.

The present invention, which does not rely on the
35 detection of the presence of double-stranded regions, is also designed to increase the accuracy of detection and quantitation of polynucleotide analytes in hybridization

assays. The invention increases both the sensitivity and specificity of such assays, by reducing the incidence of signal generation that occurs in the absence of target, and does not involve a substantial increase in either time or cost relative to current assay configurations. In certain embodiments, the invention is also effective in compensating for the loss in signal that can result when background noise is reduced.

10 Disclosure of the Invention

Methods and kits are provided for detecting nucleic acid analytes in a sample. In general, the methods represent improvements on solution phase sandwich hybridization which involve binding the analyte to a solid support, labelling the analyte, and detecting the presence of label on the support. Preferred methods involve the use of amplification multimers which enable the binding of significantly more label in the analyte-probe complex, enhancing assay sensitivity and specificity.

20 In a first aspect of the invention, an assay is provided in which two or more distinct "capture extender" molecules are used, each of which must bind to the analyte in order for the assay to result in a detectable signal. As noted above, capture extender molecules are bridging probes which bind to the analyte as well as to support bound "capture probes." In one embodiment, at least two capture extender molecules must bind to a single support-bound capture probe in order for the assay to result in a detectable signal.

30 In a further, related aspect of the invention, an assay is provided in which the melt temperature T_{m1} of the multicomponent complex formed between the analyte and support-bound capture probes, mediated by two or more distinct capture extender molecules, is significantly higher than the melt temperature T_{m2} of each two-component complex formed between a capture probe and an individual capture extender molecule. In this aspect, the assay is carried out

- 5 -

at conditions which favor formation of hybrid complexes in which analyte molecule is bound to the capture probes. This technique is premised on the enhanced stability of the multi-component complex relative to the less stable two-component
5 complexes. A preferred method of favoring analyte-bound hybrid complexes includes running one or more steps of the assay at a temperature between T_{m1} and T_{m2} .

In another aspect of the invention, an assay is provided in which two or more distinct "label extender"
10 molecules are used; as noted before, label extender molecules are bridging probes which bind to the analyte as well as to label probes, either directly, as in U.S. Patent No. 4,868,105, or indirectly through amplification multimers, as in U.S. Patent No. 5,124,246. Multiple label extenders must
15 bind to the analyte in order for a positive signal (indicating presence of the analyte in the sample) to be generated.

In another related aspect of the invention, an assay is provided in which the melt temperature T_{m1} of the
20 multicomponent complex formed between the analyte and an amplification multimer or label probe, mediated by two or more distinct label extender molecules, is significantly higher than the melt temperature T_{m2} of each two-component complex formed between an amplification multimer or label
25 probe and an individual label extender molecule. In this aspect, the assay is carried out at conditions which favor formation of hybrid complexes in which analyte molecule is bound to the amplification multimers or label probes. This technique is premised on the enhanced stability of the multi-
30 component complex relative to the much less stable two-component complexes. A preferred method of favoring analyte-amplification multimer hybrid complexes includes running one or more steps of the assay at a temperature between T_{m1} and T_{m2} .

35 In still another aspect of the invention, amplification assays are carried out with two distinct amplification multimers which are bridged by one or more

label probes. Each label probe contains two regions, each approximately 5 to 40 nucleotides in length, preferably 10 to 20 nucleotides in length, which are complementary to corresponding regions in each amplification multimer. The length of the complementary regions is selected so as to ensure that the melting temperature of the complex formed between the label probe and a single amplification multimer will be lower, preferably at least about 10°C lower than the melting temperature of the complex formed between the two amplification multimers, mediated by one or more label probes. Thus, as with the assays described above, an individual multimer will not form a stable hybrid with an individual label probe; however, multi-component hybrid complexes formed from at least one label probe and at least two multimers are stable. Since the multicomponent complex is more likely to form when the amplification multimers are placed in proximity through binding to analyte, this technique produces a more target-dependent signal.

In yet another aspect of the invention, a variation on the aforementioned assay is provided in which two distinct label probes are provided, wherein the two distinct label probes must bind together in order for a signal to be produced. As with the preceding assays, specificity is enhanced as a result of the additional probe sets and the additional hybridization steps which must take place in order for a detectable signal to be generated.

The invention also encompasses variations on the aforementioned assays, in which, for example, oligonucleotide competitors are incorporated into the assay so as to bind to the capture probes (thus reducing the likelihood of nonspecific hybridization on the solid support), and wherein shorter capture probes are used (again, to reduce the likelihood of nonspecific hybridization on the support). Oligonucleotide competitors may also be used to inhibit binding between the label extenders and the amplification multimers, or between the label probes and the amplification multimers.

Further, the invention encompasses methods for compensating for the loss in signal which can result from the various techniques provided herein for reducing background noise. These methods involve the use of preamplifier molecules which serve as intermediate moieties between label extender molecules and amplification multimers, and are structured so as to bind a plurality of amplification multimers. In this way, the number of label probes per label extender can be vastly increased.

The invention additionally encompasses a method for carrying out a hybridization assay in which each of the aforementioned techniques are combined, i.e., in which two or more distinct label extender molecules are used, two or more distinct capture extender molecules are used, amplification multimers and label probes are structured such that label probes bridge adjacent multimers, and the like.

Finally, the invention encompasses kits containing the reagents necessary to carry out the solution phase sandwich hybridization assays described and claimed herein.

Brief Description of the Figures

Figure 1 diagrams a nucleic acid hybridization assay of the prior art.

Figures 2 through 7 are specific examples of noise-producing hybridization events.

Figures 8, 9 and 10 are examples of the improved nucleic acid hybridization assay of the present invention in which capture extender molecules are required to bind a target to the solid support. Figures 8 and 9 show different ways in which two capture extenders bind to a single capture probe.

Figure 10 is an example of the improved nucleic acid hybridization assay of the present invention using multiple capture extender molecules that bind to the same or different capture probes, one per probe.

Figure 11 is an example of the improved nucleic acid hybridization assay of the present invention using label extender molecules that form a cruciform structure.

5 Figure 12 is an example of the improved nucleic acid hybridization assay of the present invention using multiple amplification multimers and bridging label probes.

Figure 13 is an example of the improved nucleic acid hybridization assay of the present invention using multiple amplification multimers and multiple bridging label
10 probes.

Figure 14 shows the melting curves of capture extenders-labeled target-capture probe and labeled capture extenders-capture probe with 2 hours equilibration.

Figure 15 is another example of the improved
15 nucleic acid hybridization assay of the present invention using multiple amplification multimers and multiple bridging label probes.

Figure 16 is an example of the improved nucleic acid hybridization assay of the present invention combining
20 several of the individual concepts disclosed herein.

Modes for Carrying Out the Invention

Definitions and nomenclature:

Before the present invention is disclosed and
25 described in detail, it is to be understood that this invention is not limited to specific assay formats, materials or reagents, as such may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only and is not
30 intended to be limiting.

In this specification and in the claims which follow, reference will be made to a number of terms which shall be defined to have the following meanings:

As used herein, the terms "polynucleotide" and
35 "oligonucleotide" shall be generic to polydeoxyribonucleotides (containing 2-deoxy-D-ribose), to polyribonucleotides (containing D-ribose), to any other type of polynucleotide

which is an N-glycoside of a purine or pyrimidine base, and to other polymers containing nonnucleotidic backbones (e.g., peptide nucleic acids (PNAs) and synthetic sequence-specific nucleic acid polymers commercially available from the Antivirals, Inc., Corvallis, Oregon, as Neugene™ polymers) or nonstandard linkages, providing that the polymers contain nucleobases in a configuration which allows for base pairing and base stacking, such as is found in DNA and RNA. There is no intended distinction in length between the term "polynucleotide" and "oligonucleotide," and these terms will be used interchangeably. These terms refer only to the primary structure of the molecule. Thus, these terms include double- and single-stranded DNA, as well as double- and single-stranded RNA and DNA:RNA hybrids, and also include known types of modifications, for example, labels which are known in the art, methylation, "caps," substitution of one or more of the naturally occurring nucleotides with an analog, internucleotide modifications such as, for example, those with uncharged linkages (e.g., methyl phosphonates, phosphotriesters, phosphoramidates, carbamates, etc.) and with charged linkages (e.g., phosphorothioates, phosphorodithioates, etc.), those containing pendant moieties, such as, for example, proteins (including nucleases, toxins, antibodies, signal peptides, poly-L-lysine, etc.), those with intercalators (e.g., acridine, psoralen, etc.), those containing chelators (e.g., metals, radioactive metals, boron, oxidative metals, etc.), those containing alkylators, those with modified linkages (e.g., alpha anomeric nucleic acids, etc.), as well as unmodified forms of the polynucleotide or oligonucleotide.

It will be appreciated that, as used herein, the terms "nucleoside" and "nucleotide" will include those moieties which contain not only the known purine and pyrimidine bases, but also other heterocyclic bases which have been modified. Such modifications include methylated purines or pyrimidines, acylated purines or pyrimidines, or other heterocycles. Modified nucleosides or nucleotides will

also include modifications on the sugar moiety, e.g., wherein one or more of the hydroxyl groups are replaced with halogen, aliphatic groups, or are functionalized as ethers, amines, or the like.

5 The term "polynucleotide analyte" refers to a single- or double-stranded nucleic acid molecule which contains a target nucleotide sequence. The analyte nucleic acids may be from a variety of sources, e.g., biological fluids or solids, food stuffs, environmental materials, etc.,
10 and may be prepared for the hybridization analysis by a variety of means, e.g., addition of proteinase K/SDS, chaotropic salts, or the like, or phenol/chloroform extraction. The term "polynucleotide analyte" is used interchangeably herein with the terms "analyte," "analyte
15 nucleic acid," "target" and "target molecule."

 As used herein, the term "target region" or "target nucleotide sequence" refers to a probe binding region contained within the target molecule. The term "target
20 sequence" refers to a sequence with which a probe will form a stable hybrid under desired conditions.

 As used herein, the term "probe" refers to a structure comprised of a polynucleotide, as defined above, which contains a nucleic acid sequence complementary to a nucleic acid sequence present in the target molecule. The
25 polynucleotide regions of probes may be composed of DNA, and/or RNA, and/or synthetic nucleotide analogs.

 It will be appreciated that the binding sequences need not have perfect complementarity to provide stable hybrids. In many situations, stable hybrids will form where
30 fewer than about 10% of the bases are mismatches, ignoring loops of four or more nucleotides. Accordingly, as used herein the term "complementary" intends to refer to an oligonucleotide that forms a stable duplex with its
"complement" under assay conditions, generally where there is
35 about 90% or greater homology.

 The terms "nucleic acid multimer" or "amplification multimer" are used herein to refer to a linear or

- 11 -

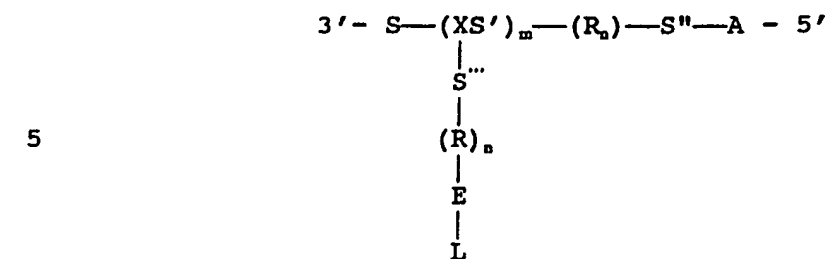
branched polymer of the same repeating single-stranded oligonucleotide segment or different single-stranded polynucleotide segments, each of which contains a region where a label probe can bind, i.e., contains a nucleic acid sequence complementary to a nucleic acid sequence contained within a label probe; the oligonucleotide segments may be composed of RNA, DNA, modified nucleotides or combinations thereof. At least one of the segments has a sequence, length, and composition that permits it to bind specifically to a target sequence in a label probe; additionally, at least one of the segments has a sequence, length, and composition that permits it to bind specifically to a target sequence in a label extender or preamplifier. Typically, such segments will contain approximately 15 to 50, preferably 15 to 30, nucleotides, and will have a GC content in the range of about 20% to about 80%. The total number of oligonucleotide segments in the multimer will usually be in the range of about 3 to 1000, more typically in the range of about 10 to 100, and most typically about 50. The oligonucleotide segments of the multimer may be covalently linked directly to each other through phosphodiester bonds or through interposed linking agents such as nucleic acid, amino acid, carbohydrate or polyol bridges, or through other cross-linking agents that are capable of cross-linking nucleic acid or modified nucleic acid strands. Alternatively, the multimer may be comprised of oligonucleotide segments which are not covalently attached, but are bonded in some other manner, e.g., through hybridization. Such a multimer is described, for example, in U.S. Patent No. 5,175,270 to Nilsen et al. The site(s) of linkage may be at the ends of the segment (in either normal, 3'-5' orientation or randomly oriented) and/or at one or more internal nucleotides in the strand. In linear multimers the individual segments are linked end-to-end to form a linear polymer. In one type of branched multimer three or more oligonucleotide segments emanate from a point of origin to form a branched structure. The point of origin may be another nucleotide segment or a multifunctional molecule to

which at least three segments can be covalently bound. In another type, there is an oligonucleotide segment backbone with one or more pendant oligonucleotide segments. These latter-type multimers are "fork-like," "comb-like" or
5 combination "fork-" and "comb-like" in structure, wherein "comb-like" multimers, the preferred multimers herein, are polynucleotides having a linear backbone with a multiplicity of sidechains extending from the backbone. The pendant
10 segments will normally depend from a modified nucleotide or other organic moiety having appropriate functional groups to which oligonucleotides may be conjugated or otherwise attached. The multimer may be totally linear, totally
15 branched, or a combination of linear and branched portions. Typically, there will be at least two branch points in the multimer, more preferably at least three, more preferably in the range of about 5 to 30, although in some embodiments there may be more. The multimer may include one or more segments of double-stranded sequences. Further information concerning multimer synthesis and specific multimer
20 structures may be found in commonly owned U.S. Patent No. 5,124,246 to Urdea et al.

PCT Publication No. WO 92/02526 describes the comb-type branched multimers which are particularly preferred in conjunction with the present method, and which are
25 composed of a linear backbone and pendant sidechains; the backbone includes a segment that provides a specific hybridization site for analyte nucleic acid or nucleic acid bound to the analyte, whereas the pendant sidechains include iterations of a segment that provide specific hybridization
30 sites for a labeled probe.

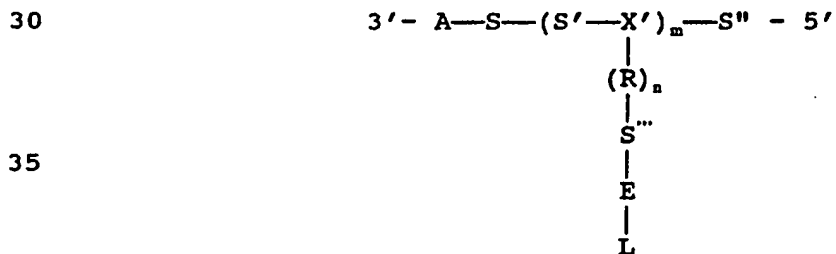
A first type of preferred comb-type polynucleotide multimer may be represented by the following schematic formula (I):

- 13 -



where S is a first spacer segment of at least about 15 nucleotides, preferably about 15 to 50 nucleotides, X is a multifunctional nucleotide that provides a branch site, S' is a branching site spacer segment of 0 to about 15 nucleotides, preferably 0 to 10 nucleotides, m is an integer equal to or greater than 15, preferably in the range of 15 to 100, R is a cleavable linker molecule, n is 0 or 1, S'' is a second spacer segment of about 0 to 10 nucleotides, preferably 5 to 10 nucleotides, A is a segment that is capable of hybridizing specifically to analyte nucleic acid or nucleic acid bound to analyte, S''' is a third spacer segment of 0 to 10 nucleotides, E is an oligonucleotide extension of 5 to 10 nucleotides and L is a segment containing 2 to 10 iterations, preferably 3 to 6 iterations, of a nucleotide sequence that is capable of hybridizing specifically to a labeled oligonucleotide probe.

A second type of preferred embodiment of these comb-type polynucleotide multimers may be represented by the following schematic formula (II):

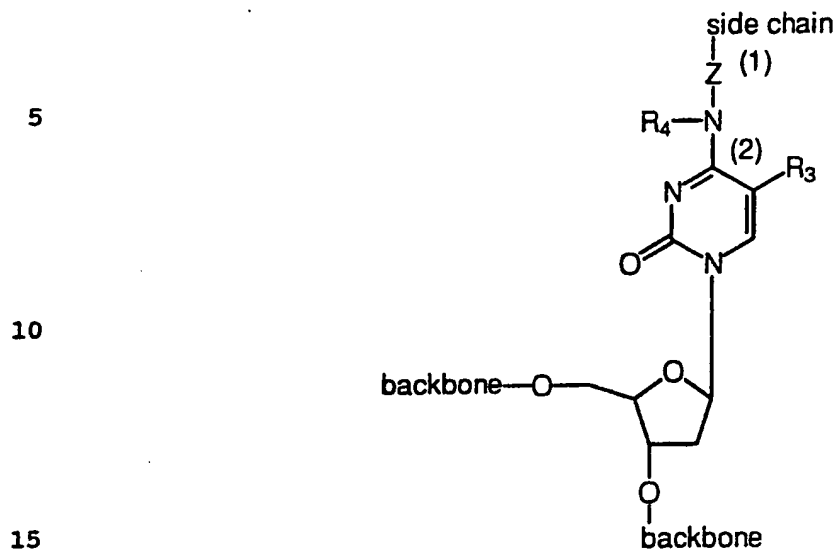


where A is a segment that is capable of hybridizing specifically to analyte nucleic acid or nucleic acid bound to analyte, S is a first spacer segment of at least about 15 molecules, preferably about 15 to 50 molecules, X' is a

monomeric molecule that provides a branch site, S' is a branching site spacer segment of 0 to about 15 molecules, preferably 0 to 10 molecules, m is an integer equal to or greater than 15, preferably in the range of 15 to 100, S" is
5 a second spacer segment of about 0 to 10 molecules, preferably 5 to 10 molecules, R is a cleavable linker molecule, n is 0 or 1, S''' is a third spacer segment of 0 to 10 molecules, E is an oligonucleotide extension of 5 to 10 nucleotides and L is a segment containing 2 to 10 iterations,
10 preferably 3 to 6 iterations, of a nucleotide sequence that is capable of hybridizing specifically to a labeled oligonucleotide probe.

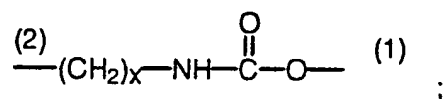
The entire backbone of the multimer or the portion thereof from S to S'', inclusive, and the portion of
15 the sidechain excluding L will typically be synthesized chemically as an integral unit using conventional automated solid-phase oligonucleotide synthesis chemistry and equipment. In this regard, the spacer segment S serves to space the portion of the molecule that contains the branching
20 sites from the solid phase (the 3' end of S is bound directly or indirectly to the surface of the solid phase). In other embodiments the entire backbone and the pendant sidechains including L may be synthesized as an integral unit.

The modified nucleotides or branching monomer
25 designated X or X' in the above formulae may be a multifunctional nucleotide in which one functional group is used for sidechain extension and the others are used for backbone bonds. Examples of multifunctional nucleotides are described in EPA 883096976 (U.S. Serial No. 340,031), the
30 disclosure of which is incorporated herein by reference. These modified nucleotides are preferably of the formula:

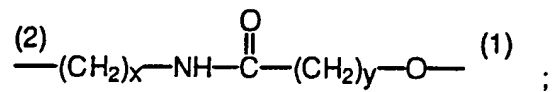


where R³ is hydrogen, methyl, I, Br or F, R⁴ is hydrogen or methyl, Z is selected from the group consisting of

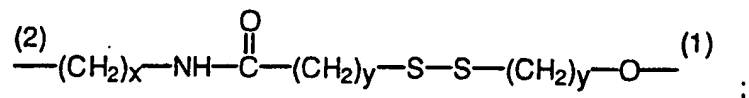
20



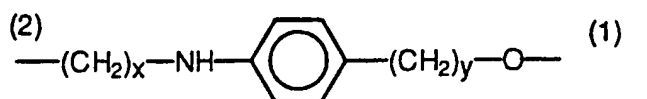
25



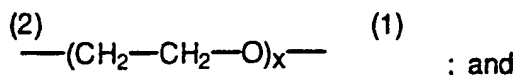
30



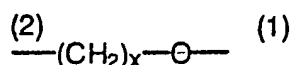
35



5



10



15 wherein x and y may be the same or different and are integers in the range of 1 to 8, inclusive. (The designations "(1)" and "(2)" at the Z linkage indicate the orientation of the Z linker moiety.)

For multimers of Formula I, as indicated, the
 20 spacer segment S' is optional and may be used, if desired, to space each branch site from preceding/ succeeding flanking branch sites or a series of adjacent branch sites from flanking series of branch sites. The second spacer segment S'' is also optional and may be employed to space the branched
 25 portion of the molecule from the segment A to which the analyte is ultimately bound (through one or more intermediate molecules such as label extenders and preamplifiers). Such spacing has been found to improve the binding between the analyte and the multimer. Likewise, the third spacer segment
 30 S''' is optional. It is preferably polyT.

For multimers of Formula II, as indicated, the
 spacer segment S' is optional and may be used, if desired, to
 space each branch site from preceding/ succeeding flanking
 branch sites or a series of adjacent branch sites from
 35 flanking series of branch sites. Likewise, the spacer segment S''' is optional. S, S', S'', and S''' may comprise nucleotidic or nonnucleotidic molecules. An example of a

nonnucleotidic molecule which may be used in a spacer segment is the cleavable linker molecule R, described below.

Segment A has a sequence and length that permits it to bind specifically and stably to a nucleic acid, such as a label extender or a preamplifier, which is bound to the analyte. In order to achieve such specificity and stability segment A will normally be 15 to 50, preferably 15 to 30, nucleotides in length. The specific length and sequence of this segment will, of course, vary depending upon the nucleic acid to which it is intended to hybridize.

Segment E is a sidechain extension that is chemically synthesized using automated solid-phase oligonucleotide synthesis equipment and techniques. It is typically about 5 to 10 nucleotides in length and serves as a site to which segment L may be ligated enzymatically or chemically.

Segment L comprises iterations of an oligomer segment that is capable of hybridizing specifically and stably to a labeled oligonucleotide probe. These segments are also typically 15 to 150, preferably 15 to 120, nucleotides in length. Each L segment will normally contain 2 to 10 iterations of the segment, preferably 3 to 6 iterations. Some sidechains may not include an L segment. Normally at least about 50% of the sidechains, preferably at least about 70% of the sidechains, will include an L segment.

The cleavable linker molecules (R) in the backbone and/or sidechains are optional, but preferred. They provide selectable cleavage sites so that samples of the large, comb-type polynucleotide may be cleaved for analysis and characterization purposes. In this regard it is preferred that there be cleavage sites in each sidechain and additional cleavage sites just 5' or the 5'-most branch site (for multimers of formula I) or where the sidechain joins the backbone (for multimers of formula II). Examples of cleavable linker molecules that may be incorporated into the polynucleotides are disclosed in EPA 883096976.

The polynucleotides of the invention may be assembled using a combination of solid phase direct oligonucleotide synthesis, enzymatic ligation methods, and solution phase chemical synthesis as described in detail in
5 PCT Publication No. WO 92/02526.

As noted above, a "preamplifier" molecule may also be used, which serves as a bridging moiety between the label extender molecules and the amplification multimers. In this way, more amplifier and thus more label is bound in any
10 given target-probe complex. Preamplifier molecules may be either linear or branched, and typically contain in the range of about 30-3000 nucleotides. In the preferred embodiment herein, the preamplifier molecule binds to at least two
15 different label extender molecules, such that the overall accuracy of the assay is increased (i.e., because, again, a plurality of hybridization events are required for the probe-target complex to form).

As used herein, a "biological sample" refers to a sample of tissue or fluid isolated from an individual,
20 including but not limited to, for example, plasma, serum, spinal fluid, semen, lymph fluid, the external sections of the skin, respiratory, intestinal, and genitourinary tracts, tears, saliva, milk, blood cells, tumors, organs, and also samples of *in vitro* cell culture constituents (including but
25 not limited to conditioned medium resulting from the growth of cells in cell culture medium, putatively virally infected cells, recombinant cells, and cell components). Preferred uses of the present method are in detecting and/or quantitating (a) viral nucleic acids, such as from hepatitis
30 B virus ("HBV"), hepatitis C virus ("HCV"), hepatitis D virus ("HDV"), human immunodeficiency virus ("HIV"), and the herpes family of viruses, including herpes zoster (chicken pox), herpes simplex virus I & II, cytomegalovirus, Epstein-Barr virus, and the recently isolated Herpes VI virus, and (b)
35 bacterial nucleic acids, such as Chlamydia, Mycobacterium tuberculosis, etc.

As used herein, the term "nonspecific hybridization" is used to refer to those occurrences in which a segment of a first polynucleotide which is intended to hybridize to a segment of a selected second polynucleotide instead hybridizes to a third polynucleotide, triggering an erroneous result, i.e., giving rise to a situation where label may be detected in the absence of target molecule.

As used herein, the term "nonspecific binding" is used to refer to those occurrences in which a polynucleotide binds to the solid support through an interaction--which may be either direct or indirect--that does not involve hybridization.

Referring now to the preferred embodiment represented in Figure 1, the following terms apply to the hybridization assay depicted therein.

"Label extender molecules (LEs)," also referred to herein as "label extenders," contain regions of complementarity vis-à-vis the analyte polynucleotide and to the amplifier multimer ("AMP"). If a preamplifier is used (not shown in the figure), the label extender molecules will bind to this intermediate species rather than directly to the amplifier multimer. If neither preamplifier nor amplifier is used, the label extender molecules will bind directly to a sequence in the label probe ("LP"). Thus, label extender molecules are single-stranded polynucleotide chains having a first nucleic acid sequence L-1 complementary to a sequence of the analyte polynucleotide, and a second region having a multimer recognition sequence L-2 complementary to a segment M-1 of label probe, amplifier multimer or preamplifier.

"Label probes (LPs)" are designed to bind either to the label extender, or, if an amplification multimer is employed in the assay, to the repeating oligonucleotide segments of the multimer. LPs either contain a label or are structured so as to bind to a label. Thus, LPs contain a nucleic acid sequence L-3 complementary to a nucleic acid sequence M-2 present within the label probe or the repeating oligonucleotide segments of the multimer and are bound to, or

- 20 -

structured so as to bind to, a label which provides, directly or indirectly, a detectable signal.

"Capture extender molecules (CEs)," also referred to herein as "capture extenders," bind to the analyte polynucleotide and to capture probes, which are in turn bound to a solid support. Thus, capture extender molecules are single-stranded polynucleotide chains having a first polynucleotide sequence region containing a nucleic acid sequence C-1 which is complementary to a sequence of the analyte, and a second, noncomplementary region having a capture probe recognition sequence C-2. The sequences C-1 and L-1 are nonidentical, noncomplementary sequences that are each complementary to physically distinct sequences of the analyte.

"Capture probes (CPs)" bind to the capture extenders and to a solid support. Thus, as illustrated in Figure 1, capture probes have a nucleic acid sequence C-3 complementary to the C-2 sequence of a CE and are covalently (or otherwise tightly) bound to a solid support.

Generally, solution phase hybridization assays carried out using the system illustrated in Figure 1 proceed as follows. Single-stranded analyte nucleic acid is incubated under hybridization conditions with the capture extenders and label extenders. The resulting product is a nucleic acid complex of the analyte polynucleotide bound to the capture extenders and to the label extenders. This complex may be subsequently added under hybridizing conditions to a solid phase having the capture probes bound to the surface thereof; however, in a preferred embodiment of this invention, the initial incubation is carried out in the presence of the support-bound capture probes. The resulting product comprises the complex bound to the solid phase via the capture extender molecules and capture probes. The solid phase with bound complex is then optionally separated from unbound materials. An amplification multimer, preferably a comb-type multimer as described above, is then optionally added to the solid phase-analyte-probe complex under

- 21 -

hybridization conditions to permit the multimer to hybridize to the LEs; if preamplifier probes are used, the solid phase-analyte-probe complex is incubated with the preamplifier probes either along with the amplification multimer or prior to incubation with the amplification multimer. The resulting solid phase complex is then separated from any unbound preamplifier and/or multimer by washing. The label probes are then added under conditions which permit hybridization to the LEs, or, if an amplification multimer was used, to the repeating oligonucleotide segments of the multimer. The resulting solid phase labeled nucleic acid complex is then washed to remove unbound labeled oligonucleotide, and remaining label is measured. It should be noted that the components represented in Figure 1 are not necessarily drawn to scale, and that the amplification multimers, if used, contain a far greater number of repeating oligonucleotide segments than shown (as explained above), each of which is designed to bind a label probe.

As will be appreciated by those skilled in the art, the techniques of the present invention may be used in conjunction with a wide variety of assay formats. However, for simplicity, the present techniques will be discussed in terms of the aforementioned solution phase hybridization assay using amplification multimers, which represents the preferred embodiment herein. Various sources of background noise which can arise in such an assay are illustrated in Figures 2 through 7. It will be noted that each of these figures depicts a situation where label will be detected on the solid support in the absence of target. In Figures 3, 4 and 5, nucleotide sequences present in the amplifier molecule, the label extender and the label probe, respectively, bind to the capture probe rather than to the desired sequences. In Figures 2, 6 and 7, the capture probe has hybridized to the capture extender molecule properly, but an incorrect molecule has then hybridized to the capture extender. In Figure 2, the amplifier has hybridized directly to the capture extender, in the absence of target molecule,

while in Figures 6 and 7, the label probe and the label extender have hybridized directly to the capture extender, again giving rise to a situation where label will be detected in the absence of analyte. It will be appreciated, however, that other such erroneous--i.e., "nonspecific"--hybridization and binding scenarios can be envisioned wherein a signal is generated in the absence of target. The techniques of the invention address these other potential sources of background noise as well.

The primary focus of the present method is on eliminating a number of sources of background noise, by maximizing the interaction of capture extender and label extender probes with the target molecule, minimizing the interaction of capture probes and capture extender molecules with the label probes, label extender molecules and amplifiers, increasing the number of probes and/or hybridization steps necessary to give rise to a target-dependent signal, and reducing the likelihood that incorrect moieties will bind to the support-bound capture probes.

In a first embodiment of the invention, a hybridization assay is provided which is configured such that the temperature T_{m1} at which the target molecule "melts" from the support-bound capture probes (defined as the temperature at which 50% of the individual capture probes participating in target molecule/capture extender/capture probe complexes are no longer bound to the target molecule) is significantly higher than the temperature T_{m2} at which an individual capture extender molecule "melts" from a single capture probe. This procedure may be used in virtually any type of hybridization assay wherein capture probes and capture extender molecules are used, including a wide range of solution phase hybridization assays, amplification assays, filter hybridization methods, assays involving the polymerase chain reaction ("PCR"), and the like. One example of a hybridization assay with which the present technique is useful is that described in U.S. Patent No. 4,868,105 to Urdea et al., or, preferably, that described above in

conjunction with the configuration illustrated in Figure 1 and described above. This method is premised on the design and construction of hybrid complexes such that the melt temperature T_{m1} at which the analyte dissociates from the capture probe in the capture probe-capture extender-analyte hybrid is at least about 5°C greater than, preferably at least about 10°C greater than the melt temperature T_{m2} at which a capture extender dissociates from a capture probe in the capture probe-capture extender hybrid.

10 This stability difference is exploited in the assay by conducting at least one step in the assay under stringency conditions which favor the T_{m1} complex formation, but disfavor the T_{m2} complex formation. Stringency can be controlled by altering a step parameter which is a
15 thermodynamic variable. Such variables are well known in the art, and include formamide concentration, salt concentration, chaotropic salt concentration, pH (hydrogen ion concentration), organic solvent content, and temperature. "Chaotropic salt" refers to a salt that acts as a hydrophobic
20 bond breaker, including the trihaloacetates, isothiocyanate and perchlorate. Hamaguchi et al., J. Am. Chem. Soc. (1962) 84:1329-1338. A preferred stringency control is temperature: at least one assay step is conducted at a temperature between T_{m1} and T_{m2} , more preferably about midway between the two
25 temperatures. A preferred step at which stringency is exercised is the initial hybridization step in the assay, in which target is incubated with capture extender molecules and support bound capture probes. Thus, in a preferred embodiment, the initial hybridization step is carried out at
30 a temperature which is higher than T_{m2} but lower than T_{m1} . Since nonspecifically hybridized molecules can bind through the capture probe and capture extender molecules (as illustrated in Figures 2, 6 and 7), this method significantly reduces certain types of nonspecific hybridization.

35 It will be readily apparent to one skilled in the art that the greater the temperature difference between T_{m1}

and T_{m2} , the greater the "efficiency" of this technique in removing background noise. Thus, one skilled in the art will recognize that temperature differentials of less than 10°C, even less than 5°C, would also permit reduction of background noise, albeit to a lesser extent. One could increase efficiency in such situations by increasing the number of steps which utilize appropriate stringency conditions, or by repeating a single stringent step.

Preferably, the method is carried out using at least two distinct capture extender molecules, each of which binds to a distinct segment of the analyte. The capture extender molecules comprise a first nucleotide sequence complementary to a segment of analyte and a second nucleotide sequence complementary to a capture probe. This assay has two primary topological embodiments.

A first embodiment of this assay configuration in which two distinct capture extender molecule are used is illustrated in Figures 8 and 9. In this embodiment, the two distinct capture extenders have distinct first nucleotide sequences complementary to distinct but proximate segments of analyte, and also have distinct second nucleotide sequences complementary to distinct segments of a single capture probe. "CE1" and "CE2" represent the two different capture extender molecules, positioned in a cruciform-like structure, such that each extender molecule hybridizes to proximate but distinct segments of target, and to proximate but distinct segments of a single capture probe.

As illustrated in Figure 9, the capture probe is structured so as to contain: (1) a first nucleotide sequence C-1 which binds to a nucleotide sequence C-3 in first capture extender CE1; and (2) a different nucleotide sequence C-2 which binds to a nucleotide sequence C-4 in second capture extender CE2. CE1 and CE2 then hybridize to distinct, nonoverlapping segments of the analyte molecule. Preferably, sequences C-1, C-2, C-3 and C-4 are relatively short, i.e., less than about 30 nucleotides in length, and preferably in the range of about 10 to 15 nucleotides in length. C-1 and

C-2 can be directly adjacent, or separated by a spacer region. In addition, it is preferred that the binding of capture probe to capture extender molecules (i.e., C-1:C-3 and C-2:C-4) be relatively weak (T_m less than about 55°), while the binding of the capture probes to the target through the capture extender molecules be much stronger (T_m greater than about 65°). This allows the target molecule to bind to the solid support with far greater stability, on the order of 100- to 1000-fold, than the capture extender molecules. This method also enables use of fewer capture probes, which in turn reduces the likelihood of nonspecific hybridization as illustrated in Figures 3, 4 and 5. This assay is exemplified in the experimental section herein, in Examples 1 and 2.

It will be appreciated by those skilled in the art that the cruciform-type configuration shown in Figure 8 is for purposes of exemplification only, and that alternative assay configurations employing two or more capture extender molecules are also possible. The only requirement is that the assay be structured such that the target binds to the solid support with a melt temperature greater than that of the capture extenders binding to the capture probe. It will also be appreciated by those skilled in the art that the embodiment of Figure 9 works equally well if C-1 and C-2 are identical capture probe sequences complementary to identical sequences C-3 and C-4 in the two capture extenders: in this instance, the capture probe contains two copies of the repeat sequence C-1.

A second embodiment of this assay configuration in which two distinct capture extender molecule are used is illustrated in Figure 10. In this embodiment, two or more (preferably more than three) distinct capture extenders have distinct first nucleotide sequences complementary to distinct but proximate segments of analyte, and also have second nucleotide sequences complementary to a segment of a capture probe. In this embodiment, only one capture extender binds per capture probe. If all the capture probes in the assay contain a single sequence that binds to the capture

- 26 -

extenders, then the capture extenders will all have the same second nucleotide sequence complementary to the binding segment of the capture probe. Alternatively, the assay may use distinct capture probes containing multiple sequences that bind to the capture extenders, in which case the capture extenders will have distinct second nucleotide sequences.

Thus, in Figure 10, "CE3", "CE4" and "CE5" represent three different capture extenders, such that each capture extender contains distinct first nucleotide sequences that hybridize to distinct segments of target. CE3 and CE4 contain a second nucleotide sequence C-6 which binds to a nucleotide sequence C-5 in a first capture probe CP1; while CE5 contains a different second nucleotide sequence C-8 which binds to a nucleotide sequence C-7 in second capture probe CP2. It will be easily seen that any combination of two or more such capture extenders can be used in this assay.

Another variation on this technique involves the use of two or more distinct label extender molecules, each of which must bind to the target in order for the amplifier probe to bind. This assay is carried out basically as described above with respect to Figure 1; however, as noted, at least two distinct label extender molecules are incorporated into the assay. The label extenders comprise a first nucleotide sequence complementary to a segment of analyte and a second nucleotide sequence complementary to an amplification multimer (or preamplifier, as discussed below).

In this assay configuration in which two distinct label extenders are used is illustrated in Figure 11. In this embodiment, the two distinct label extenders have distinct first nucleotide sequences complementary to distinct but proximate segments of analyte, and also have distinct second nucleotide sequences complementary to distinct segments of a single amplification multimer. "LE1" and "LE2" represent the two different label extenders positioned in a cruciform-like structure, such that each extender molecule hybridizes to proximate but distinct segments of target, and to proximate but distinct segments of a single amplification

- 27 -

multimer. This assay is exemplified in the experimental section herein, in Example 2.

As illustrated in Figure 11, the amplification multimer is structured so as to contain: (1) a first nucleotide sequence C-1 which binds to a nucleotide sequence C-3 in first label extender LE1; and (2) a different nucleotide sequence C-2 which binds to a nucleotide sequence C-4 in second label extender LE2. LE1 and LE2 then hybridize to distinct, nonoverlapping segments of the analyte molecule. Preferably, sequences C-1, C-2, C-3 and C-4 are relatively short, i.e., less than about 30 nucleotides in length, and preferably in the range of about 10 to 15 nucleotides in length. C-1 and C-2 can be directly adjacent, or separated by a spacer region. In addition, it is preferred that the binding of amplification multimer to label extender molecules (i.e., C-1:C-3 and C-2:C-4) be relatively weak (T_m less than about 45°), while the binding of the amplification multimer to the target through the label extender molecules be much stronger (T_m greater than about 65°). This allows the target molecule to bind to the amplification multimer with far greater stability, on the order of 100- to 1000-fold, than the label extender molecules. Again, as with the preceding method in which at least two capture extender molecules are used, assay specificity is increased by virtue of the additional hybridization steps which are necessary to give rise to a target-dependent signal.

It will be appreciated by those skilled in the art that the cruciform-type configurations shown in Figure 11 is for purposes of exemplification only, and that alternative assay configurations employing two or more label extender molecules are also possible. The only requirement is that the assay be structured such that the target binds to the amplification multimer with a melt temperature greater than that of the label extenders binding to the amplification multimer. It will also be appreciated by those skilled in the art that this embodiment works equally well if C-1 and C-2 are identical amplification multimer sequences

complementary to identical sequences C-3 and C-4 in the two label extenders: in this instance, the amplification multimer contains two copies of the repeat sequence C-1. It will further be appreciated that the preceeding description
5 could be utilized with a preamplifier (as discussed below), wherein the label extenders interact in cruciform structure with the preamplifier rather than directly with the amplification multimer.

In another embodiment of the invention, the
10 phenomenon of target-independent signal generation is addressed by bridging adjacent amplifier molecules in such a way as to reduce virtually all of the principal sources of assay background, including nonspecific hybridization of label extender molecules to capture probes and capture
15 extender molecules, nonspecific hybridization of amplification multimers to capture probes and capture extender molecules and amplifier nonspecific binding. In this embodiment, two distinct amplifier multimers are used, designated AMP1 and AMP2 in Figure 12, as well as two
20 distinct label extender molecules, designated LE1 and LE2. Neither AMP1 nor AMP2 will retain label unless they are within bridging distance of each other, so that there is a much higher likelihood that the amplifiers are actually bound to the target molecule before labelling occurs. This is
25 accomplished by providing label probes which contain: (1) a first nucleic acid sequence L-1 which contains a nucleic acid sequence complementary to a region in the repeating oligonucleotide subunits of AMP1; (2) a second nucleic acid sequence L-2 which contains a nucleic acid sequence
30 complementary to a region in the repeating oligonucleotide subunits of AMP2; and (3) a detectable label therebetween. L-1, L-2, and the corresponding complementary sequences in the amplifier probes are selected such that the melting temperature of the complex formed from both amplifier probes
35 and the label probes is preferably at least about 10°C higher than the melting temperature of the complex formed between the label probe and a single amplifier multimer. It will

also be appreciated that such a configuration gives rise to the advantages discussed above with respect to the use of multiple probes and consequent additional hybridization steps required to produce a detectable signal.

5 In a variation on this embodiment, two or more distinct label probes may be used, either containing two different types of labels, as shown in Figure 13, or containing an inactive segment of a label which is then activated upon conjugation with a label probe on an adjacent
10 amplifier molecule. Examples of this multi-amplifier, multi-label probe embodiment include those wherein adjacent label probes contain: (a) individual subunits, members or portions of an enzyme such as β -galactosidase; (b) an enzyme and a corresponding co-enzyme (such as a flavoenzyme and FADH or an
15 NAD-linked dehydrogenase and NADH); or (c) enzymes that form a part of a channelling system or cascade system wherein the product of one enzyme is the substrate for the second enzyme (such as the fatty-acid synthetase system). In each case the two label probes contain distinct moieties which do not
20 produce the detectable product unless brought together by the presence of the target and thus amplifier bridging.

 In a variation on all of the above embodiments, preamplifier molecules may be used in all of the above embodiments to increase signal. Preamplifiers are added to
25 the assay reaction either concurrently with or prior to the addition of the amplification multimers.

 As alluded to earlier, a primary cause of background noise, i.e., signals which are produced and detected independent of target molecule, results from the
30 support-bound capture probes and the fact that polynucleotides other than capture extender molecules can bind thereto. Additional embodiments of the invention derive from this realization. First, solid supports used in hybridization assays like those described above are
35 configured such that the capture probes are shorter, typically less than about 20 nt, and more typically about 15 nt. Second, oligonucleotides containing sequences which are

- 30 -

identical to C-2 as illustrated in Figure 1 are introduced into the assay; such oligonucleotides function as competitors for the capture probes and thus reduce the number of available capture probe hybridization sites (C-3 in Figure 1). This is illustrated in Example 1.

Experimental

The practice of the present invention will employ, unless otherwise indicated, conventional techniques of synthetic organic chemistry, biochemistry, molecular biology, and the like, which are within the skill of the art. Such techniques are explained fully in the literature. See, e.g., Sambrook, Fritsch & Maniatis, Molecular Cloning: A Laboratory Manual, Second Edition (1989); Oligonucleotide Synthesis (M.J. Gait, ed., 1984); Nucleic Acid Hybridization (B.D. Hames & S.J. Higgins, eds., 1984); and a series, Methods in Enzymology (Academic Press, Inc.). All patents, patent applications, and publications mentioned herein, both *supra* and *infra*, are hereby incorporated by reference.

It is to be understood that while the invention has been described in conjunction with the preferred specific embodiments thereof, that the description above as well as the examples which follow are intended to illustrate and not limit the scope of the invention. Other aspects, advantages and modifications within the scope of the invention will be apparent to those skilled in the art to which the invention pertains.

In the following examples, efforts have been made to insure accuracy with respect to numbers used (e.g., amounts, temperature, etc.) but some experimental error and deviation should be accounted for. Unless indicated otherwise, temperature is in degrees C and pressure is at or near atmospheric.

Example 1

Amplification Assay Using Different Capture Extenders in a "Cruciform" Format

- 31 -

This example describes a hybridization assay using two or more distinct capture extenders, as illustrated in Figure 8. The goal of this experimental work was to reduce background signals caused by capture extender probe binding to the solid support. By reducing the ability of capture extender probes to bind to the support, the target polynucleotide is forced to bind through multiple capture extender probes in order to be bound stably to the solid surface, giving rise to an assay background which is as low as in an assay run with no capture extender probes (i.e., because essentially no capture extender molecules would be bound to the support). The experimental work summarized in this example also demonstrates that when molecules capable of competitively binding to the immobilized capture probe are added, it is possible to reduce the background even further, because the likelihood that the label extender molecules or label probes will bind to the support via the immobilized capture probe is significantly reduced.

A capture probe designated CP2 was attached to the solid support. Two pairs of HCV capture extender molecules were designed. The first member of each pair had a 5' sequence complementary to the last 16 nts of capture probe CP2 and the second member of each pair had a 3' sequence complementary to the first 13 nts of CP2. The extenders could thus bind to the solid support as illustrated in Figure 8. They can bridge neighboring CP2 molecules (as the target in Figure 10, which has been captured via CEs 3, 4 and 5 to two different CP molecules) or they can form a cruciform with a single CP2 (as the target in Figure 9, which has bound via CEs 1 and 2 to the same CP molecule). Since the probes are designed optimally to form the cruciform and since the capture probe concentration is kept low, the cruciform should be greatly favored kinetically over the bridging of two neighboring CP2 molecules and the cruciform is presumed to predominate in the equilibrium distribution.

A signal amplification solution phase nucleic acid sandwich hybridization assay format was employed in this

- 32 -

example. The signal amplification is generated through a branched DNA multimer (amplifier) which has a first segment (E') that hybridizes to a segment of the label extender probes and up to fifteen iterations of a segment (F), wherein
5 segment F hybridizes to three labeled oligonucleotides. The target nucleic acid is bound to the solid support via an immobilized capture probe and a capture extender probe which hybridizes to both the capture probe and the target. The amplifier multimer is bound to the immobilized target via a
10 label extender probe which hybridizes to both the target and the amplifier multimer. Two competitor probes were also used for background reduction. These probes bind to the capture probe.

The capture extender probes, label extender
15 probes and competitor probes as used in this assay were as follows.

Sequence (5'-->3') Capture extender probes (the segment which binds to the immobilized capture probe is underlined):

20

1: (SEQ ID NO.: 1)

TTTCAGCAATCAGGTGTTCTCGTCCTGGCAATCCGGTGTACTCACCGGTTT

2: (SEQ ID NO.: 2)

TTTCAGCAATCAGGTGTTCTCWTTCGGCGATTCCGGTGTACTCACCGGTTT

25 3: (SEQ ID NO.: 3)

GTATTGAGCGGGTTKMTCCAAGAAAGGACCCGGTCTGGGCTCTGGGAC

4: (SEQ ID NO.: 4)

GCATAGAGTGGGTTWATCCAAGAAAGGACCCAGTCTGGGCTCTGGGAC

5: (SEQ ID NO.: 5)

30 TTTCAGCAATCAGGTGTTTCAGCAGTCTTGCGGGGGCAGCCCCAAATCTCCAG

6: (SEQ ID NO.: 6)

TTTCAGCAATCAGGTGTTTCAGTGATCTTGCGGGGGCGTGCCCCAAATCTCCAG

7: (SEQ ID NO.: 7)

TTTCAGCAATCAGGTGTTTCAGCAGTCTCGCGGGGGCAGCCCCAAATCTCCAG

35 8: (SEQ ID NO.: 8)

TTTCAGCAATCAGGTGTTTCAGCAGTCTTGCGGGGGCAGCCCCAAATGGCTGG

9: (SEQ ID NO.: 9)

- 33 -

TTTCAGCAATCAGGTGTTTCAGTGATCTCGCGGGGGCACGCCCAAATTTCTGG

10: (SEQ ID NO.: 10)

ACAAGGCCTTTTCGCGACCCAACACTACTCGGCTTCGGCTCTGGGAC

11: (SEQ ID NO.: 11)

5 ACAAGGCCKTTTCGCAACCCAACGCTACTMGGCTTCGGCTCTGGGAC

Label extender probes used (the sequence which hybridizes to the amplifier multimer is underlined):

10 12: (SEQ ID NO.: 12)

AGGCATAGGACCCGTGTCTTTCTCACAGGGGAGTGATTCATGGTGGAGTGTC

13: (SEQ ID NO.: 13)

AGGCATAGGACCCGTGTCTTATGGCTAGGCGCTTTCTGCGTGAAGACAGTAGT

14: (SEQ ID NO.: 14)

15 AGGCATAGGACCCGTGTCTTKCCTGGAGGCTGTACGASACTSGTACTAGCGCC

15: (SEQ ID NO.: 15)

AGGCATAGGACCCGTGTCTTCGCAGACCACTATGGCTCTCCCGGGAGGGGGGG

16: (SEQ ID NO.: 16)

AGGCATAGGACCCGTGTCTTGGGGCACTCGCAAGCACCCCTATCAGGCAGTACC

20 17: (SEQ ID NO.: 17)

AGGCATAGGACCCGTGTCTTTGTGCTCATGKTGCACGGTCTACGAGACCTCCC

Competitor probes (these sequences hybridize to the immobilized capture probe):

25

18: (SEQ ID NO.: 18)

TCGGCTCTGGGAC

19: (SEQ ID NO.: 19)

CAGCAATCAGGTGTTC

30

The plates used for the assay were coated as follows: White Microlite 1 removawell strips (polystyrene microtiter plates, 96 wells/plate) were purchased from Dynatech Inc.

35

Each well was filled with 250 μ l 1N HCl and incubated at room temperature for 15-20 min. The plates were then washed 1 times with 1 x PBS and the wells aspirated to

- 34 -

remove liquid. The wells were then filled with 250 μ l 1N NaOH and incubated at room temperature for 15-20 min. The plates were again washed 3 times with 1 x PBS and the wells aspirated to remove liquid.

5 Poly(phe-lys) was purchased from Sigma Chemicals, Inc. This polypeptide has a 1:1 molar ratio of phe:lys and an average m.w. of 47,900 gm/mole. It has an average length of 309 amino acids and contains 155 amines/molecule. The polypeptide was mixed with 2M NaCl/1 x PBS to a final
10 concentration of 0.1 mg/mL (pH 6.0). 200 μ l of this solution was added to each well. The plate was wrapped in plastic to prevent drying and incubated at 30°C overnight. The plate was then washed 3 times with 1 x PBS and the wells aspirated to remove liquid.

15 To 250 OD₂₆₀ units/ml of the following oligonucleotide (Capture Probe CP2):
5'-XGTCCAGAGCCGAGAACACCTGATTGCTG-3' (SEQ ID NO.: 20)
(X is the long chain amine modified nucleotide (N⁴-
(6-aminocaproyl-2-aminoethyl) derivative of
20 5-methylcytidine)) in 50 mM sodium phosphate pH 7.8 was added 180 mg of bis(sulfosuccinimidyl) suberate (BS³). The mixture was vortexed and incubated at room temperature for 30 min. A gel filtration column (Pharmacia Sephadex G-25, NAP-25) equilibrated with 10 mM sodium phosphate pH 6.5 was used to
25 purify the activated oligonucleotide. The activated oligonucleotide reaction mixture was applied to the column and allowed to filter. The eluate was collected and saved for use in the next step. The concentration of the eluate was adjusted to 3.7×10^{-2} OD₂₆₀ units/ml using 50 mM sodium
30 phosphate pH 7.8 for dilution.

100 μ l of the activated oligonucleotide-containing eluent was added to each well and the wells were incubated at 4°C for 12-18 hours. The plate was then washed 2 times with 1 x PBS and the wells aspirated to remove
35 liquid.

250 μ l of 0.2N NaOH containing 0.1 wt.% SDS was added to each well. The plate was wrapped in plastic and

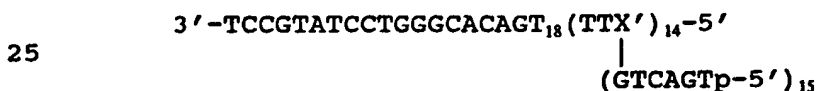
- 35 -

incubated at 65°C for 60 min. The plate was then washed 3 times with 1 x PBS and the wells aspirated to remove liquid.

100 µl of 50 mM sodium phosphate pH 7.8 with 0.4 mg/ml BS³ was to each well and allowed to incubate with the plate for 12- 18 hours. The plate was then washed 2 times with 1 x PBS and 1 time with water. The plates were stored in plastic containers with desiccant at 4°C. The amplifier multimer was prepared as follows:

All chemical syntheses of oligonucleotides were performed on an automatic DNA synthesizer (Applied Biosystems, Inc., (ABI) model 380 B). Phosphoramidite chemistry of the β-cyanoethyl type was used including 5'-phosphorylation which employed PHOSTEL™ reagent (DMT-O-CH₂CH₂-(SO₂)-CH₂CH₂-O-P(N(iPr)₂)(-O-CH₂CH₂CN) wherein DMT is dimethoxytrityl and iPr is isopropyl). Standard ABI protocols were used except as indicated. Where it is indicated that a multiple of a cycle was used (e.g., 1.2 cycle), the multiple of the standard amount of amidite recommended by ABI was employed in the specified cycle.

A comb body of the following structure was first prepared:

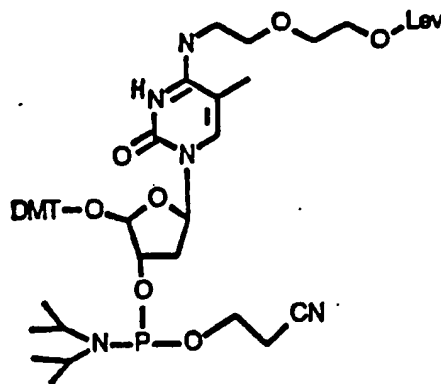


wherein X' is a branching monomer and p is a phosphate.

The portion of the comb body through the 14 (TTX') repeats was first synthesized using 33.8 mg aminopropyl-derivatized thymidine controlled pore glass (CPG) (2000 Å, 7.4 micromoles thymidine per gram support) with a 1.2 cycle protocol. The branching site nucleotide was of the formula:

5

10



For synthesis of the comb body (not including sidechain extensions), the concentration of β -cyano-ethylphosphoramidite monomers was 0.1 M for A, C, G and T, and 0.15 M for the branching site monomer X'. Detritylation was done with 3% trichloroacetic acid in methylene chloride using stepped flowthrough for the duration of the deprotection.

A six-base sidechain extension of the formula 3'-GTCAGTp was synthesized at each branching monomer site as follows. Two sidechain extensions were synthesized onto the terminal X' branching site. To remove the levulinyl group, a solution of 0.5 M hydrazine hydrate in pyridine/glacial acetic acid (1:1 v/v) was introduced and kept in contact with the CPG support for 90 min with renewal of the liquid every 15 min, followed by extensive washing with pyridine/glacial acetic acid (1:1 v/v) and then by acetonitrile. After the deprotection the six base sidechain extensions were added using a 6.4 cycle.

In these syntheses the concentration of phosphoramidites was 0.1 M (except 0.2 M Phostel™ phosphorylating reagent).

Detritylation was effected with a solution of 3% trichloroacetic acid in methylene chloride using continuous flowthrough, followed by a rinse solution of toluene/chloromethane (1:1 v/v). Branched polynucleotide

- 37 -

toluene/chloromethane (1:1 v/v). Branched polynucleotide chains were removed from the solid supports automatically in the DNA synthesizer. The ammonium hydroxide solution was collected in 4 ml screw-capped Wheaton vials and heated at 5 60°C for 12 hr to remove all base-protecting groups. After cooling to room temperature the solvent was removed in a Speed-Vac evaporator and the residue dissolved in 100 µl water. The sidechains and a ligation template/linker of the following structures were also made using the automatic 10 synthesizer:

Sidechain:

3'-GATGCG(TTCATGCTGTTGGTGTAG)₃-5'(p) (SEQ ID NO.: 21)

15 Ligation template for linking sidechain extension:

3'-CGCATCACTGAC-5'(p) (SEQ ID NO.: 22)

The crude comb body was purified by a standard polyacrylamide gel (7% with 7 M urea and 1X TBE running 20 buffer) method.

The sidechains were ligated to the comb body as follows. The comb body (4 pmole/µl), sidechains (93.75 pmole/µl), and sidechain linking template (75 pmoles/µl) were combined in 1 mM ATP/ 5 mM DTT/ 50 mM Tris-HCl, pH 8.0/ 10 mM 25 MgCl₂/ 2 mM spermidine. The mixture was heated in a water bath to 95°C, and then slowly cooled to below 35°C over a 1 hr period. The concentrations of the components of the mixture were adjusted to 2 mM ATP, 10 mM DTT, 14% polyethylene glycol, then 0.2 units/µl T4 DNA ligase were 30 added,. The mixture was incubated for 16-24 hr at 23°C. The DNA was precipitated in NaCl/ethanol and resuspended in water. Ligation products were then purified by polyacrylamide gel electrophoresis.

35 The hybridization assay was carried out as follows:

- 38 -

As a target, a synthetic 615 nucleotide transcript was prepared using a pGEM (Promega) vector containing nucleotides 54-668 of the HCV J1 clone and SP6 RNA polymerase. The negative control was no target.

5 Sample preparation consisted of delivering 200 μ l of 2 mg/ml proteinase K in 0.07 M Tris-HCl, pH 8.0/0.7 M LiCl/0.06 M sodium phosphate/0.06 M EDTA, pH 7.0/0.7% SDS/50 fmoles capture extender probes (each)/200 fmole label extender probes (each)/5000 fmole competitor probes (each))
10 to each well. Different control experiments contained various combinations of the probes as indicated in the results table below. Plates were agitated to mix the contents in the well, covered and incubated for 16 hr at 63°C.

15 After a further 10 minute period at room temperature, the contents of each well were aspirated to remove all fluid, and the wells washed 2X with washing buffer (0.1% SDS/0.015 M NaCl/ 0.0015 M sodium citrate). The amplifier multimer was then added to each well (50 μ l of 0.7
20 fmole/ μ l solution in 0.48 M NaCl/0.048 M sodium citrate/0.1% SDS/0.5% diethyl pyrocarbonate treated "blocking reagent" (a purified fraction of dry milk powder, Boehringer Mannheim, catalog No. 1096 176). After covering the plates and agitating to mix the contents in the wells, the plates were
25 incubated for 30 min at 45°C.

 After a further one minute period at room temperature, the wells were washed as described above.

 Alkaline phosphatase label probe, disclosed in EP 883096976, was then added to each well (50 μ l/well of 2.66
30 fmoles/ μ l). After incubation at 45°C for 15 min, and 1 min at room temperature, the wells were washed three times as above and then three times with 0.015 M NaCl/0.0015 M sodium citrate.

 An enzyme-triggered dioxetane (Schaap et al.,
35 Tet. Lett. 28:1159-1162 (1987) and EPA Pub. No. 0254051), obtained from Lumigen, Inc., was employed. 50 μ l Lumiphos 530 (Lumigen) was added to each well. The wells were tapped

lightly so that the reagent would fall to the bottom and gently swirled to distribute the reagent evenly over the bottom. The wells were covered and incubated at 37°C for 20-40 min.

5 Plates were then read on a Dynatech ML 1000 luminometer. Output was given as the full integral of the light produced during the measurement period of 250 ms.

Table 1

10	Line	Capture Extender	Competitor	Target	Relative Light Units (Std Dev)
	1	No	No	No	0.47 (0.08)
	2	Yes	No	No	0.47 (0.10)
15	3	Yes	Yes	No	0.34 (0.01)
	4	Yes	No	Yes	38.0 (2.2)
20	5	Yes	Yes	Yes	40.2 (7.1)

With non-cruciform extenders, typically the "no capture extender" control has about 2-fold lower background than the control sample containing capture extenders. This is because molecules (LEs, AMP, and label) bind to the CEs. However, a comparison of line 1 and line 2 shows that the background with cruciform capture extenders is the same as the background without cruciform capture extenders.

30 Comparison of lines 1, 2 and 3 shows that with competitor it is possible to reduce backgrounds below the level of the no capture extender control. Presumably this is due to blocking label extender binding to the capture probe. Comparison of lines 4 and 5 shows that competitors do not compete out

35 target binding. Target binding is not affected by competitors because target is bound much more tightly via multiple CEs, which is much tighter than the binding of individual extender probes or individual competitors.

Example 2Amplification Assay Using Multiple
Label Extenders in a "Cruciform" Format

This example describes a hybridization assay using two different label extenders, as illustrated in Figure 11. The goal of this experimental work was to reduce background signals caused by label extenders binding to the solid support or to the capture extender or capture probe molecules, i.e., the type of background which is generated when a label extender binds to the support in a manner not mediated by the target. Once the label extender is bound, the preamplifier, the amplifier multimer and the label probes (in this experiment, alkaline phosphatase label probes were used) can bind and generate signal that is not dependent on the presence of target. By reducing the ability of label extender probes to bind to the amplifier multimer individually, the preamplifier is forced to bind through multiple label extender probes in order to be bound stably to the surface. A preamplifier would not remain bound to a single label extender during the hybridization conditions of the assay. The label extenders are designed in a way where the two label extenders necessary to facilitate the binding of the amplifier multimer are located proximal to each other when they are bound to the target sequence. A single hybridized label extender will not efficiently bind preamplifier under the reaction conditions. In this way, the binding of the preamplifier is only favorable in the presence of target.

A synthetic oligonucleotide capture probe called PSCP (see Sequence #41) was attached to the plates. A set of HCV label extender probes were designed. Each label extender contains a sequence (T) complementary to the target in addition to one or two additional sequences (chosen from sequences A, B, C, or D) which bind to a preamplifier probe. This preamplifier contains sequences (A' and B', or C' and D') which bind to the LEs and eight repeats of a sequence (E') which binds the amplifier multimer.

- 41 -

A signal amplification solution phase nucleic acid sandwich hybridization assay format was employed in this example. The signal amplification is generated through a branched DNA multimer (amplifier) which is designed to have a first segment (E) that hybridizes to a segment of the preamplifier (E') and up to fifteen iterations of a segment (F), wherein segment F hybridizes to up to three alkaline phosphatase-labeled oligonucleotides. The target nucleic acid is bound to the solid support via capture probes and a plurality of capture extenders which hybridize to both the capture probes and the target. The capture extender probes, label extender probes and preamplifier probes as used in this assay were as follows.

Sequence (5'-->3') Capture extender probes (the segment which binds to the immobilized capture probe is underlined):

20: (SEQ ID NO.: 23)

TCCTCACAGGGGAGTGATTCATGGTGGAGTGTCTCTTGGAAGAAAGTGAAGTG

21: (SEQ ID NO.: 24)

ATGGCTAGACGCTTTCTGCGTGAAGACAGTAGTCTCTTGGAAGAAAGTGAAGTG

22: (SEQ ID NO.: 25)

TCCTGGAGGCTGCACGACRCTCATACTAACGCCCTCTTGGAAGAAAGTGAAGTG

23: (SEQ ID NO.: 26)

25 CGCAGACCACTATGGCTCTYCCGGGAGGGGGGCTCTCTTGGAAGAAAGTGAAGTG

24: (SEQ ID NO.: 27)

TCRTCCYGGCAATTCGGGTGTA CTACCGGTTCTCTTGGAAGAAAGTGAAGTG

Label extender probes used (the sequence which hybridizes to the target is underlined, the sequence as abbreviated above are enclosed in parentheses):

25: (A-T) (SEQ ID NO.: 28)

CTGAGTTTCTGGCTCGCATTGAGCGGGTTDATCCAAGAAAGGACCCGG

26: (B-T-C) (SEQ ID NO.: 29)

ACTGAGTCAGTCAGTCAGCAGTCTYGGGGGGCACGCCCAARTCTCCAGGAAAGTTTGAAT
ATG

5 27: (A-T-D) (SEQ ID NO.: 30)

CTGAGTTTCTGGCTCACAAGGCCTTTCGCAACCCAACACTACTCGGCTACCTACCTACCTA
CCT

28: (B-T-C) (SEQ ID NO.: 31)

AGTCAGTCAGTCAGTCGGGGCACTCGCAAGCACCCATCAGGCAGTACCGAAAGTTTGAAT

10 ATG

29: (A-T-D) (SEQ ID NO.: 32)

CTGAGTTTCTGGCTCYGTGCTCATGRTGCACGGTCTACGAGACCTCCACCTACCTACCTA
CCT

30: (B-T-C) (SEQ ID NO.: 33)

15 AGTCAGTCAGTCAGTCGTTACGTTTGKTTYTTTGTGGTTTTRGGAWTGAAAGTTTGAAT
ATG

31: (T-D) (SEQ ID NO.: 34)

CGGGAACCTTRACGTCCTGTGGGCGRCGGTTGGTACCTACCTACCTACCT

20 Label extender probes used for the control experiment were
(the sequence which hybridizes to the target is underlined):

:HCV.33.13 (SEQ ID NO.: 35)

AGGCATAGGACCCGTGTCTTCARGTAAACTCCACCRACGATCTGRCCCRCC

25

:HCV.33.14 (SEQ ID NO.: 36)

AGGCATAGGACCCGTGTCTTRCGCACACCCAAYCTRGGGGCCCTGCGCGGCAA

:HCV.33.15 (SEQ ID NO.: 37)

30 AGGCATAGGACCCGTGTCTTAGGTTGCGACCGCTCGGAAGTCTTYCTRGTCGC

- :HCV.33.16A (SEQ ID NO.: 38)
AGGCATAGGACCCGTGTCTTTCGHRCCCTTGGGGATAGGCTGACGTCWACCTCG
- :HCV.33.16B (SEQ ID NO.: 39)
5 AGGCATAGGACCCGTGTCTTTCGHRCCCTTGGGGATAGGTTGTGCGCCWTCCACG
- :HCV.33.17 (SEQ ID NO.: 40)
AGGCATAGGACCCGTGTCTTTCGRCGGCTGRGCCCAGRYCCTRCCCTCGGRYYG
- 10 :HCV.33.18 (SEQ ID NO.: 41)
AGGCATAGGACCCGTGTCTTTCBHRCCCTCCTTTRCCRTAGAGGGGCCADGGRTA
- :HCV.33.19 (SEQ ID NO.: 42)
AGGCATAGGACCCGTGTCTTTCGRCGGGGWGACAGGAGCCATCCYGCCACCC
15
- :HCV.33.20 (SEQ ID NO.: 43)
AGGCATAGGACCCGTGTCTTTCGCGGGGTCYGTGGGGCCCCAYCTAGGCCGRGA
- :HCV.33.21 (SEQ ID NO.: 44)
20 AGGCATAGGACCCGTGTCTTTCGATGACCTTACCCAARTTRCGCGACCTRCG
- :HCV.33.22 (SEQ ID NO.: 45)
AGGCATAGGACCCGTGTCTTTCGCCATGAGRTC GGCGAAGCCGCAYGTRAGGGT
- 25 32: (SEQ ID NO.: 46)
AGGCATAGGACCCGTGTCTTTCGATTGAGCGGGTTDATCCAAGAAAGGACCCGG
- 33: (SEQ ID NO.: 47)
AGGCATAGGACCCGTGTCTTTCGAGTCTYCGGGGGCAGCCCCAARTCTCCAG
30
- 34: (SEQ ID NO.: 48)
AGGCATAGGACCCGTGTCTTTCGAAGGCCTTTCGCAACCCAACACTACTCGGCT
- 35: (SEQ ID NO.: 49)
35 AGGCATAGGACCCGTGTCTTTCGGGCACTCGCAAGCACCTATCAGGCAGTACC
- 36: (SEQ ID NO.: 50)

- 44 -

AGGCATAGGACCCGTGTCTTYGTGCTCATGRTGCACGGTCTACGAGACCTCCC

37: (SEQ ID NO.: 51)

AGGCATAGGACCCGTGTCTTGTTACGTTTGKTTYTTYTTTGRGGTTTRGGAWT

5

38: (SEQ ID NO.: 52)

AGGCATAGGACCCGTGTCTTCGGGAACTTRACGTCCTGTGGGCGRCGGTTGGT

10 Preamplifier probes used (sequence E' is underlined, the starting and ending points for A', B', C', and D' are indicated by lowercase letters, the remaining sequences are for spacing):

39: (SEQ ID NO.: 53)

15 AGGCATAGGACCCGTGTCTTTTTTAGGCATAGGACCCGTGTCTTTTTTAGGCATAGGACCC
GTGTCCGTGGATGTTTGAGGCATAGGACCCGTGTCTTTTTTAGGCATAGGACCCGTGTCTT
TTTTAGGCATAGGACCCGTGTGCGTAGTGACTGAGGCATAGGACCCGTGTCTTTTTTAGG
CATAGGACCCGTGTCTTTTTTb'-CATATTCAAACCTTTC-b'a'-GAGCCAGAACTCAGT
-a'

20

40: (SEQ ID NO.: 54)

AGGCATAGGACCCGTGTCTTTTTTAGGCATAGGACCCGTGTCTTTTTTAGGCATAGGACCC
GTGTCCGTGGATGTTTGAGGCATAGGACCCGTGTCTTTTTTAGGCATAGGACCCGTGTCTT
TTTTAGGCATAGGACCCGTGTGCGTAGTGACTGAGGCATAGGACCCGTGTCTTTTTTAGG
25 CATAGGACCCGTGTCTTTTTTd'-AGGTAGGTAGGTAGGT-d'
c'-GACTGACTGACTGACT-c'

30 The plates used for the assay were coated as described above except instead of the DNA sequence CP2, the sequence PSCP was attached to the plate.

PSCP:

41: (SEQ ID NO.: 55)

5'- XCA CTT CAC TTT CTT TCC AAG AG-3'

35

- 45 -

The amplifier multimer was prepared as described above. The hybridization assay was carried out as follows:

A standard curve of HCV was prepared by diluting a high titer hepatitis C virus sample in HCV negative human serum and delivering 50 μ l aliquots of dilutions corresponding to a range of 1,500 to 19,500 viral equivalents to wells of microtiter dishes prepared as described above.

Sample preparation consisted of delivering 150 μ l P-K Buffer (3.3 mg/ml proteinase K in 58 mM Tris-HCl, pH 8.0/0.6 M NaCl/0.06 M sodium citrate/12 mM EDTA, pH 8.0/1.3%SDS/16 μ g/ml sonicated salmon sperm DNA/7% formamide/100 fmoles capture extender probes/400 fmoles label extender probes) to each well. The control experiment used 50 fmol/well capture extender probes, 160 fmol/well label extender probes in the same buffer. Plates were agitated to mix the contents in the well, covered and incubated for 16 hr at 63°C. The control experiment was left at 63°C during the "preamplification" step.

After a further 10 minute period at room temperature, the contents of each well were aspirated to remove all fluid, and the wells washed 2X with washing buffer (0.1% SDS/0.015 M NaCl/ 0.0015 M sodium citrate). The preamplifier probes were then added (100 fmole in 50 μ l 0.75 M NaCl/0.075 M sodium citrate/0.1% SDS/0.5% blocking reagent (as above, Boehringer Mannheim, catalog No. 1096 176)) to each well (except control). Plates were agitated, covered and incubated at 53°C for 30 min.

The plates were then cooled and washed as described above. The amplifier multimer was then added to each well (85 fmol in same buffer used for preamplifier probes). The control experiment used 30 fmol/well amplifier multimer in the same buffer. After covering the plates and agitating to mix the contents in the wells, the plates were incubated for 30 min at 53°C.

After a further 10 min period at room temperature, the wells were washed as described above.

Alkaline phosphatase label probe, disclosed in EP 883096976, was then added to each well (125 fmol in 50 μ l 0.75 M NaCl/0.075 M sodium citrate/0.1% SDS/0.5% blocking reagent (as above, Boehringer Mannheim, catalog No. 1096 176)). After incubation at 53°C for 15 min, and 10 min at room temperature, the wells were washed twice as above and then 3X with 0.015 M NaCl/0.0015 M sodium citrate.

An enzyme-triggered dioxetane (Schaap et al., Tet. Lett. (1987) 28:1159-1162 and EPA Pub. No. 0254051), obtained from Lumigen, Inc., was used as the luminescence substrate. 50 μ l Lumiphos 530 (Lumigen) was added to each well. The wells were covered, agitated for 30 sec and incubated at 37°C for 25 min.

Plates were then read on a Dynatech ML 1000 luminometer. Output was given as the full integral of the light produced during the 250 millisec read time.

Results are shown in the Table below.

	Target Concentration	With Preamplifiers (Relative Light Units)	Without Preamplifiers (Relative Light Units)
20	1	3.87 (\pm 0.26)	1.58 (\pm 0.15)
	2	1.64 (\pm 0.15)	0.80 (\pm 0.09)
25	3	1.08 (\pm 0.16)	0.61 (\pm 0.09)
	0	0.63 (\pm 0.05)	0.43 (\pm 0.04)

30

The assay run with preamplifier probes had a higher value for signal minus noise for each of the analyte concentrations compared to the assay run without preamplifier probes.

Example 3

Multidentate Capture

In this example, T_m measurements were carried out on 40 microwells containing the capture probe xt1* (see PCT Publication No. WO 92/02526 cited earlier herein, and EPA

- 47 -

883096976). Capture extenders and the xt1* capture probe were as follows:

5 xt1-tailed HCV capture extenders (5'-->3'; functional domains of the probes are separated by periods):

:HCV.33.1.XT1: (SEQ ID NO.: 56)
TCCTCACAGGGGAGTGATTCATGGTGGAGTGTC.CTTCTTTGGAGAAAGTGGTG

10 :HCV.33.2.XT1: (SEQ ID NO.: 57)
ATGGCTAGACGCTTCTGCGTGAAGACAGTAGT.CTTCTTTGGAGAAAGTGGTG

:HCV.33.3.XT1: (SEQ ID NO.: 58)
GCCTGGAGGCTGCACGRCACTCATACTAACGCC.CTTCTTTGGAGAAAGTGGTG
15

:HCV.33.4.XT1: (SEQ ID NO.: 59)
CGCAGACCACTATGGCTCTYCCGGGAGGGGGGG.CTTCTTTGGAGAAAGTGGTG

:HCV.33.5.XT1: (SEQ ID NO.: 60)
20 TCRTCCYGGCAATTCCGGGTGTACTCACCGGTTC.CTTCTTTGGAGAAAGTGGTG

xt1* capture probe: (SEQ ID NO.: 61)

CACCACTTTCTCCAAAGAAG
25

Capture extender probes and HCV target RNA were labeled with P-32. For the capture extender-capture probe T_m determination, labeled capture extenders were equilibrated with xt1* wells at various temperatures or in the presence of
30 various levels of the denaturant formamide. Either two hours or overnight equilibration was used. For the capture extenders-target-capture probe T_m determination, the capture extenders were hybridized to P-32 labeled target. The capture extenders-target complex was then equilibrated with
35 the capture probe wells either two hours or overnight at the various temperatures. Samples were drawn off the microwells at temperature and the percent binding determined. The T_m or

C_m were determined as the midpoints of the curves with percent binding plotted against temperature or percent formamide. To convert C_m (formamide) to T_m (no formamide), the fact that each percent formamide lowers the T_m by 0.7 degrees was used.

The melting curves of capture extenders-labeled target-xt1* capture probe and labeled capture extenders-xt1* capture probe with 2 hours equilibration are shown in Figure 14. The melting temperatures in 20% formamide and 0.6 M LiCl are: 50 degrees for capture extenders-xt1* and 60 degrees for capture extenders-target-xt1*. These numbers correspond to T_m values of 64 and 74 degrees in the absence of formamide. Figure 14 shows that 36% of the target and only 2% of the capture extenders are bound at 55 degrees in 20% formamide. This is an 18x difference that is based on target capture through multiple capture extenders.

One skilled in the art will appreciate that the difference in T_m between the target-capture extender-capture probe complex and capture extender-capture probe complex can be increased beyond 10 degrees. By lowering the capture extender-capture probe T_m , one will magnify the effect of multidentate binding.

Example 4

Hybridization Assay Using Two Amplifiers

From the previous examples it is clear that probes can be designed such that multiple (two or more) interactions are required to achieve a stable complex under the conditions of the assay. The term multiple interactions is not intended to be limiting in any way. Two types of multiple interactions have been illustrated: multiple probes forming a cruciform (4-way branched) junction and multiple capture extenders simultaneously binding to multiple capture probes on a solid support, resulting in the Target-solid support T_m being 10-20 degrees higher than the T_m of a single capture extender-capture probe hybrid.

The same concepts are extended to multiple amplifiers. A workable probe design is shown in Figure 15. In this instance two branched DNA amplifiers are bound to two label extenders. Amp1 is bound to label extender LE1 through the stable hybrid sequence X. Amp2 is bound to label extender LE2 through stable hybrid sequence Y. LE1 and LE2 do not have to be contiguous, but both must be bound to non-overlapping regions of the same target molecule. AMP1 contains branching sequences B and F and amp 2 contains branching sequences D and G. In this example, an enzyme-labeled probe contains sequences Bc and Dc, while an activator probe contains sequences Fc and Gc. In Figure 15 label extenders form stable hybrids with target and with their respective amplifiers. All other hybrids shown (involving B, D, F, G) are designed to be too weak to form stable hybrids at the temperature of the hybridization. The weak hybrids are designed to have melting temperatures about 10-20 degrees below the assay hybridization temperature. A design with melting temperatures 5 degrees below the assay temperature is also acceptable. The preferable difference is 10-20 degrees since this can increase target specific binding by 100-1000 fold.

In this way, for example, when an enzyme-labeled probe binds to sequence B, it quickly dissociates unless sequence D is nearby as well. Once a probe binds to both AMP1 and AMP2, the conformation becomes more fixed in space, kinetically favoring the next binding event. The activator probe binds independently of the labeled probe but in like manner. The binding of the activator probe is kinetically but not thermodynamically favored by the binding of the labeled probe to AMP1 and AMP2 and vice versa.

The activator probe(s) can take many forms. They can (1) activate the enzymes directly or (2) activate the enzymes indirectly by relieving inhibition or (3) they can activate the detection of the product of the enzyme reaction. As an example of method (3), the enzyme in the enzyme-labeled probe could be alkaline phosphatase and the activators could be a

- 50 -

fluoresceinated polymer similar in composition and structure to that disclosed by Schaap et al., in Clinical Chemistry 35:1863-1864 (1989). In the presence of this polymer the light output from the dephosphorylated dioxetane increased 400-fold. Ideally the luminescence activators would sandwich the enzyme-labeled probes so that dioxetane released by the enzyme would have a high probability of transferring energy to the fluorescein or other luminescent enhancers for efficient target-dependent light emission. The conditions chosen for the detection step would favor a very short lifetime of the dioxetane, reducing the efficiency of detection of dioxetane in the absence of enhancer-labeled target. By removing the fluorescein polymers or other luminescence enhancers from the bulk phosphorylated dioxetane substrate mixture, the signal will be made much more target-dependent. Even an alkaline phosphatase bound nonspecifically to the solid support that cleaves a phosphorylated dioxetane will most likely not be detected unless it survives long enough to diffuse to the target. By this method even background is converted into a type of signal (i.e. the presence of target will be required to see most of the noise).

The activator probes could also be enzyme-activators or moieties that relieve enzyme inhibition. The enzyme could have two or more subunits. Ideally it would be engineered so that the natural affinity of the subunits would be reduced or removed. In place of the amino acids responsible for the natural association would be very short oligonucleotide sequences (sequence H on one subunit and the complementary sequence Hc on the other subunit) that cannot form a stable hybrid by themselves but which would be capable of associating the two inactive subunits and thus forming an active complex if the subunits were held close to one another by virtue of a target molecule's binding both subunits to different branches of two different amplifiers as in Figure 15.

An example of activation by means of relieving enzyme inhibition would be as follows. The labeled oligonucleotide would be covalently bound to both alkaline phosphatase and theophylline, or other non-competitive enzyme inhibitor. The
5 activator probe would contain an anti-theophylline antibody with a K_a much higher than the $1/K_i$ of theophylline for alkaline phosphatase. The oligo probe containing the anti-theophylline antibody would be added in the presence of sufficient reversible denaturant such that the activator will
10 not bind to the enzyme inhibitor. Examples of such reversible denaturants include formamide and urea, which alkaline phosphatase tolerates very well. The wash steps would remove the denaturant along with the excess activator and labeled probes. At which time the target-bound anti-
15 theophylline antibody, by virtue of its proximity, would bind theophylline because $K_a \gg 1/K_i$ and thus relieve the inhibition of the alkaline phosphatase. Solid-support bound alkaline phosphatase would remain inhibited by the theophylline tethered to the labeled probe because non-specifically bound
20 anti-theophylline antibody is not close enough to relieve the inhibition.

A three-part system may also be constructed, in which branched DNA amplifiers with at least three short sequences such that two enzyme subunits (whose natural affinity for
25 each other has been reduced or removed) are brought into proximity and they are in turn surrounded by luminescent activators. In addition, a branched DNA amplifier bringing one subunit of an enzyme and another branched DNA amplifier (bound to another label extender) bringing in the other
30 subunit or an activator can be used. In addition it is equally possible to use linear amplifiers containing many different consecutive sequences to support bridging molecules. Such linear amplifiers can be easily made by ligation as discussed earlier.

35

Example 5

A Hybridization Assay Combining the Concepts

The goal of the present invention is to reduce the background noise in hybridization assays. By drastically reducing the binding of capture extenders to the solid support, the background due to molecules that bind the capture extenders is reduced. By binding an amplifier through two label extenders, background is reduced because a label extender binding to either the solid support or to any molecule bound to the solid support (including capture probe and capture extenders) is not efficiently labeled.

Similarly, an amplifier that binds to the support in the absence of target is not efficiently labeled if it forms a hybrid with the labeled probe having a T_m 10-20 degrees below the assay temperature. Also, a labeled probe that binds to the solid support or any molecule bound to the solid support is inefficiently detected if that labeled probe is not activated by its cofactors or other subunits. Activators of luminescence can increase light output more than one hundred fold (Schaap et al., supra). If the activator is removed from the substrate solution and instead bound to the target, there is a significant opportunity for noise reduction in the hybridization assay.

The present example describes how to use these concepts in a single assay. Use of any one concept results in a detectable decrease in background. Use of all of them can in principle make background undetectable, permitting the use of stronger signal amplification to increase sensitivity. A workable probe design combining these concepts is shown in Figure 16. Capture extenders and label extenders can be designed to hybridize simultaneously to the target in cruciforms. The T_m of the target binding region of the extenders is designed to be more than about 10 degrees more stable than that of hybrids involving sequences V and W and X and Y. The T_m of the capture probe--target complex is designed to be about 10 degrees or more stable than the capture extender--capture probe complex. During the hybridization a vast molar excess of the sequences 'V' and 'W' may be included to further reduce capture extender

binding to the solid support (especially in case a cool down step is included after hybridization). As shown previously, the 'V' and 'W' sequences have no effect on the target binding.

- 5 After capture is complete, an optional washing cycle is done to remove excess probes. Then, as illustrated in Figure 16, enzyme-labeled probes containing short sequences Bc and Dc and an optional activator probe containing sequences Fc and Gc are added and
- 10 hybridization conducted at a temperature approximately 20 degrees higher than the T_m s of hybrids involving sequences B, D, F, G. For example, the T_m s of sequences B, D, F, and G could be designed to be 35 degrees, while the hybridization temperature could be 50 degrees and the T_m s of the complexes
- 15 formed with AMP1 and AMP2 could be 55 to 60 degrees. Alternatively, the hybridization could be done at approximately 37 degrees in the presence of sufficient protein denaturant to (1) reduce or effectively prevent association of activator and enzyme and (2) to reduce or
- 20 effectively prevent binding of labeled probe and/or activator to just one amplifier (that is, to increase the effective hybridization temperature to approximately 50 degrees). Preferably,
- 25 protein engineering would be used to minimize interactions between enzyme and enzyme activator(s). Finally, the solid support is washed and detection is done. The preferred substrate would be a phosphorylated dioxetane. No enhancer would be present in this substrate mix, since ideally the enhancers are bound to the target. That is, either the
- 30 activator of Figure 16 is itself a luminescence activator or luminescence activators are included in the hybridization with the enzyme activators and enzyme-labeled oligonucleotide probes.

Claims

We claim:

- 5 1. In a solution phase sandwich hybridization assay for detecting a nucleic acid analyte in a sample, comprising: (a) binding the analyte directly or indirectly to a solid support; (b) labelling the analyte; and (c) detecting the presence of analyte-associated label,
- 10 the improvement which comprises incorporating a first capture extender molecule and a distinct second capture extender molecule into the assay, said first and second capture extender molecules each comprising a polynucleotide containing an analyte-binding segment capable of hybridizing
- 15 to a nucleic acid sequence present in the analyte and a support-binding segment capable of hybridizing to a nucleic acid sequence present within a capture probe bound to a solid support,
- wherein the analyte-binding segment of the first
- 20 capture extender molecule is distinct from the analyte-binding segment of the second capture extender molecule, and further wherein the capture probe contains a first capture extender binding sequence capable of hybridizing to the support-binding segment of the first capture extender
- 25 molecule, and a second capture extender binding sequence capable of hybridizing to the support-binding segment of the second capture extender molecule, such that two distinct capture extender molecules can bind to a single capture
- probe.
- 30 2. The assay of claim 1, wherein the melt temperature T_{m1} at which the nucleic acid analyte dissociates from the capture probe in the hybrid complex formed between the capture probe, capture extender molecules and nucleic acid
- 35 analyte is at least about 5°C greater than the melt temperature T_{m2} of hybrid complexes formed between the capture probe and capture extender molecules.

3. The assay of claim 2, wherein at least one step is carried out under stringency conditions which favor the T_{m1} complex formation, but disfavor the T_{m2} complex formation.

5 4. The assay of claim 3 wherein stringency is controlled by controlling at least one step parameter selected from the group consisting of formamide concentration, chaotropic salt concentration, salt concentration, pH (hydrogen ion concentration), organic
10 solvent content, and temperature.

5. The assay of claim 4 wherein the at least one step is carried out at a temperature greater than T_{m2} and less than T_{m1} .

15

6. The assay of claim 3 wherein the at least one step comprises step (a).

7. A solution phase sandwich hybridization assay for
20 detecting the presence of a nucleic acid analyte in a sample, comprising:

(a) providing a solid support having capture probes thereon, capture extender molecules having a first segment C-1 capable of hybridizing to a nucleic acid sequence in the
25 analyte and a second segment C-2 capable of hybridizing to a nucleic acid sequence in the capture probes, label extender molecules having a first segment L-1 capable of hybridizing to a nucleic acid sequence in the analyte and a second segment L-2 capable of hybridizing to a label probe system, a
30 label probe system comprising a nucleic acid sequence M-1 capable of hybridizing to L-2 and which, directly or indirectly, gives rise to a detectable signal;

(b) incubating the nucleic acid analyte under hybridization conditions with the capture extender molecules,
35 label extender molecules, and the capture probes on the solid support, simultaneously or sequentially in any order, to form a support-bound hybrid complex;

- 56 -

(c) thereafter optionally separating materials not bound to the solid support;

(d) thereafter contacting the support-bound hybrid complex with the label probe system under hybridizing conditions, to produce a labeled, support-bound hybrid complex; and

(e) thereafter separating materials not bound to the solid support; and

(f) detecting the presence of label in the labeled support-bound hybrid complex,

wherein the melt temperature T_{m1} at which the nucleic acid analyte dissociates from the capture probe in the hybrid complex formed between the capture probe, capture extender molecules and nucleic acid analyte is at least about 5°C greater than the melt temperature T_{m2} of hybrid complexes formed between the capture probe and capture extender molecules.

8. The assay of claim 7, wherein the label probe system comprises (i) an amplification multimer containing the nucleic acid sequence M-1 and a plurality of identical oligonucleotide subunits containing nucleic acid sequences M-2 capable of hybridizing to label probes, and (ii) label probes containing a nucleic acid sequence L-3 which is capable of hybridizing to M-2 and which, directly or indirectly, gives rise to a detectable signal, and wherein step (d) of the assay comprises the following steps:

(d₁) contacting the support-bound hybrid complex under hybridization conditions with the amplification multimer, to produce a second support-bound hybrid complex;

(d₂) thereafter optionally separating materials not bound to the solid support; and

(d₃) thereafter contacting the second support-bound hybrid complex with the label probes under hybridization conditions, to produce a labeled support-bound hybrid complex.

9. A solution phase sandwich hybridization assay for detecting the presence of a nucleic acid analyte in a sample, comprising:

(a) providing a solid support having capture probes thereon, capture extender molecules having a first segment C-1 capable of hybridizing to a nucleic acid sequence in the analyte and a second segment C-2 capable of hybridizing to a nucleic acid sequence in the capture probes, label extender molecules having a first segment L-1 capable of hybridizing to a nucleic acid sequence in the analyte and a second segment L-2 capable of hybridizing to a nucleic acid sequence P-1 in a preamplifier probe, a preamplifier probe having the nucleic acid sequence P-1 and capable of binding a plurality of amplification multimers through nucleic acid sequences P-2, amplification multimers containing a nucleic acid sequence M-1 capable of hybridizing to P-2 and a plurality of identical oligonucleotide subunits containing nucleic acid sequences M-2 capable of hybridizing to label probes, and label probes containing a sequence L-3 which is capable of hybridizing to M-2 and which, directly or indirectly, gives rise to a detectable signal;

(b) incubating the nucleic acid analyte under hybridization conditions with the capture extender molecules, label extender molecules, and the capture probes on the solid support, simultaneously or sequentially in any order, to form a support-bound, first hybrid complex;

(c) thereafter optionally separating materials not bound to the solid support;

(d) contacting the support-bound, first hybrid complex under hybridization conditions with the preamplifier probe and the amplification multimer, to produce a support-bound, second hybrid complex;

(e) thereafter optionally separating materials not bound to the solid support;

(f) contacting the support-bound, second hybrid complex with the label probes under hybridizing conditions, to produce a labeled, support-bound, third hybrid complex;

(g) thereafter separating materials not bound to the solid support; and

(h) detecting the presence of label in the support-bound, third hybrid complex,

5 wherein the melt temperature T_{m1} at which the nucleic acid analyte dissociates from the capture probe in the hybrid complex formed between the capture probe, capture extender molecules and nucleic acid analyte is at least about 5°C greater than the melt temperature T_{m2} of hybrid complexes
10 formed between the capture probe and capture extender molecules.

10. The assay of claim 7, wherein at least one step is carried out under stringency conditions which favor the T_{m1}
15 complex formation, but disfavor the T_{m2} complex formation.

11. The assay of claim 10 wherein stringency is controlled by controlling at least one step parameter selected from the group consisting of formamide
20 concentration, chaotropic salt concentration, salt concentration, pH (hydrogen ion concentration), organic solvent content, and temperature.

12. The assay of claim 11 wherein the at least one
25 step is carried out at a temperature greater than T_{m2} and less than T_{m1} .

13. The assay of claim 10 wherein the at least one step comprises step (b).
30

14. A solution phase sandwich hybridization assay for detecting the presence of a nucleic acid analyte in a sample, comprising:

35 (a) providing a solid support having capture probes thereon, a first capture extender molecule comprising a polynucleotide containing an analyte-binding segment capable of hybridizing to a nucleic acid sequence present in the

analyte and a support-binding segment capable of hybridizing to a nucleic acid sequence present within the capture probes, a distinct second capture extender molecule comprising a polynucleotide containing an analyte-binding segment capable of hybridizing to a nucleic acid sequence present in the analyte and a support-binding segment capable of hybridizing to a nucleic acid sequence present within the capture probes, label extender molecules having a first segment L-1 capable of hybridizing to a nucleic acid sequence in the analyte and a second segment L-2, an amplification multimer containing a nucleic acid sequence M-1 capable of hybridizing to L-2 and a plurality of identical oligonucleotide subunits containing nucleic acid sequences M-2, and label probes containing a sequence L-3 which is capable of hybridizing to M-2 and which, directly or indirectly, gives rise to a detectable signal;

(b) incubating the nucleic acid analyte under hybridization conditions with the capture extender molecules, label extender molecules, and the capture probes on the solid support, simultaneously or sequentially in any order, to form a support-bound, first hybrid complex;

(c) thereafter optionally separating materials not bound to the solid support;

(d) contacting the support-bound, first hybrid complex under hybridization conditions with the amplification multimer, to produce a support-bound, second hybrid complex;

(e) thereafter optionally separating materials not bound to the solid support;

(f) contacting the support-bound, second hybrid complex with the label probes under hybridizing conditions, to produce a labeled, support-bound, third hybrid complex; and

(g) thereafter separating materials not bound to the solid support; and

(h) detecting the presence of label in the support-bound, third hybrid complex,

wherein the capture extender molecules are configured such that the support-bound, second hybrid complex of step

- 60 -

(b) will form only when both first and second capture extender molecules have bound to the analyte.

5 15. A solution phase sandwich hybridization assay for detecting the presence of a nucleic acid analyte in a sample, comprising:

(a) providing a solid support having capture probes thereon, a first capture extender molecule comprising a polynucleotide containing an analyte-binding segment capable
10 of hybridizing to a nucleic acid sequence present in the analyte and a support-binding segment capable of hybridizing to a nucleic acid sequence present within the capture probes, a distinct second capture extender molecule comprising a polynucleotide containing an analyte-binding segment capable
15 of hybridizing to a nucleic acid sequence present in the analyte and a support-binding segment capable of hybridizing to a nucleic acid sequence present within the capture probes, label extender molecules having a first segment L-1 capable of hybridizing to a nucleic acid sequence in the analyte and
20 a second segment L-2 capable of hybridizing to a nucleic acid sequence P-1 in a preamplifier probe, a preamplifier probe having the nucleic acid sequence P-1 and capable of binding a plurality of amplification multimers through nucleic acid sequences P-2, amplification multimers containing a nucleic
25 acid sequence M-1 capable of hybridizing to P-2 and a plurality of identical oligonucleotide subunits containing nucleic acid sequences M-2 capable of hybridizing to label probes, and label probes containing a sequence L-3 which is capable of hybridizing to M-2 and which, directly or
30 indirectly, gives rise to a detectable signal;

(b) incubating the nucleic acid analyte under hybridization conditions with the capture extender molecules, label extender molecules, and the capture probes on the solid support, simultaneously or sequentially in any order, to form
35 a support-bound, first hybrid complex;

(c) thereafter optionally separating materials not bound to the solid support;

- 61 -

(d) contacting the support-bound, first hybrid complex under hybridization conditions with the preamplifier probe and the amplification multimer, to produce a support-bound, second hybrid complex;

5 (e) thereafter optionally separating materials not bound to the solid support;

(f) contacting the support-bound, second hybrid complex with the label probes under hybridizing conditions, to produce a labeled, support-bound, third hybrid complex; and

10 (g) thereafter separating materials not bound to the solid support; and

(h) detecting the presence of label in the support-bound, third hybrid complex,

wherein the capture extender molecules are configured
15 such that the support-bound, first hybrid complex of step (b) will form only when both first and second capture extender molecules have bound to the analyte.

16. The assay of claim 14, wherein the capture probe
20 contains a first capture extender binding sequence capable of hybridizing to the support-binding segment of the first capture extender molecule, and a second capture extender binding sequence capable of hybridizing to the support-binding segment of the second capture extender molecule, such
25 that two capture extender molecules can bind to a single capture probe.

17. The assay of claim 15, wherein the capture probe
contains a first capture extender binding sequence capable of
30 hybridizing to the support-binding segment of the first capture extender molecule, and a second capture extender binding sequence capable of hybridizing to the support-binding segment of the second capture extender molecule, such
that two capture extender molecules can bind to a single
35 capture probe.

- 62 -

18. A solution phase sandwich hybridization assay for detecting the presence of a nucleic acid analyte in a sample, comprising:

(a) providing a solid support having capture probes
5 thereon, capture extender molecules comprising polynucleotides containing an analyte-binding segment capable of hybridizing to a nucleic acid sequence present in the analyte and a support-binding segment capable of hybridizing to a nucleic acid sequence present within the capture probes,
10 a first label extender molecule having a first segment capable of hybridizing to a nucleic acid sequence in the analyte and a second segment capable of hybridizing to a label probe system, a distinct second label extender molecule having a first segment capable of hybridizing to a nucleic
15 acid sequence in the analyte and a second segment capable of hybridizing to a label probe system, a label probe system comprising nucleic acid sequences capable of hybridizing to the second segments of the first label extender molecule and the second label extender molecule, and which, directly or
20 indirectly, gives rise to a detectable signal;

(b) incubating the nucleic acid analyte under hybridization conditions with the capture extender molecules, label extender molecules, and the capture probes on the solid support, simultaneously or sequentially in any order, to form
25 a support-bound hybrid complex;

(c) thereafter optionally separating materials not bound to the solid support;

(d) thereafter contacting the support-bound hybrid complex with the label probe system under hybridizing
30 conditions, to produce a labeled, support-bound hybrid complex; and

(e) thereafter separating materials not bound to the solid support; and

(f) detecting the presence of label in the labeled
35 support-bound hybrid complex,

wherein the label extender molecules are configured such that the labeled support-bound hybrid complex of step

(d) will form only when both first and second label extender molecules have bound to the analyte.

19. The assay of claim 18, wherein the label probe
5 system comprises (i) an amplification multimer containing the nucleic acid sequences capable of hybridizing to the second segments of the first label extender molecule and the second label extender molecule, and a plurality of identical
oligonucleotide subunits containing nucleic acid sequences M-
10 2 capable of hybridizing to label probes, and (ii) label probes containing a nucleic acid sequence L-3 which is capable of hybridizing to M-2 and which, directly or indirectly, gives rise to a detectable signal, and wherein
step (d) of the assay comprises the following steps:
15 (d₁) contacting the support-bound hybrid complex under hybridization conditions with the amplification multimer, to produce a second support-bound hybrid complex;
(d₂) thereafter optionally separating materials not bound to the solid support; and
20 (d₃) thereafter contacting the second support-bound hybrid complex with the label probes under hybridization conditions, to produce a labeled support-bound hybrid complex.

25 20. A solution phase sandwich hybridization assay for detecting the presence of a nucleic acid analyte in a sample, comprising:

(a) providing a solid support having capture probes thereon, capture extender molecules comprising
30 polynucleotides containing an analyte-binding segment capable of hybridizing to a nucleic acid sequence present in the analyte and a support-binding segment capable of hybridizing to a nucleic acid sequence present within the capture probes,
a first label extender molecule having a first segment
35 capable of hybridizing to a nucleic acid sequence in the analyte and a second segment capable of hybridizing to a preamplifier probe, which in turn is capable of binding a

- 64 -

plurality of amplification multimers, a distinct second label
extender molecule having a first segment capable of
hybridizing to a nucleic acid sequence in the analyte and a
second segment capable of hybridizing to a preamplifier
5 probe, which in turn is capable of binding a plurality of
amplification multimers, amplification multimers containing a
nucleic acid sequence M-1 capable of hybridizing to the label
extender molecules and containing a plurality of identical
oligonucleotide subunits containing nucleic acid sequences M-
10 2, and label probes containing a sequence which is capable of
hybridizing to M-2 and which, directly or indirectly, gives
rise to a detectable signal;

(b) incubating the nucleic acid analyte under
hybridization conditions with the capture extender molecules,
15 label extender molecules, and the capture probes on the solid
support, simultaneously or sequentially in any order, to form
a support-bound, first hybrid complex;

(c) thereafter optionally separating materials not
bound to the solid support;

20 (d) contacting the support-bound, first hybrid complex
under hybridization conditions with the preamplifier probe
and the amplification multimer, to produce a support-bound,
second hybrid complex;

(e) thereafter optionally separating materials not
25 bound to the solid support;

(f) contacting the support-bound, second hybrid complex
with the label probes under hybridizing conditions, to
produce a labeled, support-bound, third hybrid complex; and

(g) thereafter separating materials not bound to the
30 solid support; and

(h) detecting the presence of label in the support-
bound, third hybrid complex,

wherein the label extender molecules are configured
such that the support-bound, second hybrid complex of step
35 (d) will form only when both first and second label extender
molecules have bound to the analyte.

- 65 -

21. A solution phase sandwich hybridization assay for detecting the presence of a nucleic acid analyte in a sample, comprising:

(a) providing a solid support having capture probes thereon, capture extender molecules having a first segment C-1 capable of hybridizing to a nucleic acid sequence in the analyte and a second segment C-2 capable of hybridizing to a nucleic acid sequence in the capture probes, a first label extender molecule having a first segment L-1 capable of hybridizing to a nucleic acid sequence in the analyte and a second segment L-2 capable of hybridizing to a first amplification multimer, a second label extender molecule having a first segment L-1a capable of hybridizing to a nucleic acid sequence in the analyte and a second segment L-2a capable of hybridizing to a second amplification multimer, a first amplification multimer containing a nucleic acid sequence M-1 capable of hybridizing to L-2 and a plurality of identical oligonucleotide subunits containing nucleic acid sequences M-2 capable of hybridizing to label probes, a second amplification multimer containing a nucleic acid sequence M-1a capable of hybridizing to L-2a and a plurality of identical oligonucleotide subunits containing nucleic acid sequences M-2a capable of hybridizing to label probes, and label probes capable of hybridizing to the amplification multimers and which, directly or indirectly, gives rise to a detectable signal;

(b) incubating the nucleic acid analyte under hybridization conditions with the capture extender molecules, label extender molecules, and the capture probes on the solid support, simultaneously or sequentially in any order, to form a support-bound, first hybrid complex;

(c) thereafter optionally separating materials not bound to the solid support;

(d) contacting the support-bound, first hybrid complex under hybridization conditions with the amplification multimers, to produce a support-bound, second hybrid complex;

- 66 -

(e) thereafter optionally separating materials not bound to the solid support;

(f) contacting the support-bound, second hybrid complex with the label probes under hybridizing conditions, to
5 produce a labeled, support-bound, third hybrid complex;

(g) thereafter separating materials not bound to the solid support; and

(h) detecting the presence of label in the support-bound, third hybrid complex,

10 wherein the label probes contain a first nucleic acid segment which is capable of hybridizing to the first amplification multimer, and a second nucleic acid segment which is capable of hybridizing to the second amplification multimer, such that the support-bound, third hybrid complex
15 is not formed unless the first and second amplification multimers are linked through the label probes.

22. A solution phase sandwich hybridization assay for detecting the presence of a nucleic acid analyte in a sample,
20 comprising:

(a) providing a solid support having capture probes thereon, capture extender molecules having a first segment C-1 capable of hybridizing to a nucleic acid sequence in the analyte and a second segment C-2 capable of hybridizing to a
25 nucleic acid sequence in the capture probes, a first label extender molecule having a first segment L-1 capable of hybridizing to a nucleic acid sequence in the analyte and a second segment L-2 capable of hybridizing to a first amplification multimer, a second label extender molecule
30 having a first segment L-1a capable of hybridizing to a nucleic acid sequence in the analyte and a second segment L-2a capable of hybridizing to a second amplification multimer, a first amplification multimer containing a nucleic acid sequence M-1 capable of hybridizing to L-2 and a plurality of
35 identical oligonucleotide subunits containing nucleic acid sequences M-2 capable of hybridizing to a first label probe segment, a second amplification multimer containing a nucleic

- 67 -

acid sequence M-1a capable of hybridizing to L-2a and a plurality of identical oligonucleotide subunits containing nucleic acid sequences M-2a capable of hybridizing to a second label probe segment, and first and second label probe segments capable of hybridizing to the amplification multimers and which, when present on proximate amplification multimers, give rise to a detectable signal;

(b) incubating the nucleic acid analyte under hybridization conditions with the capture extender molecules, label extender molecules, and the capture probes on the solid support, simultaneously or sequentially in any order, to form a support-bound, first hybrid complex;

(c) thereafter optionally separating materials not bound to the solid support;

(d) contacting the support-bound, first hybrid complex under hybridization conditions with the amplification multimer, to produce a support-bound, second hybrid complex;

(e) thereafter optionally separating materials not bound to the solid support;

(f) contacting the support-bound, second hybrid complex with the label probes under hybridizing conditions, to produce a labeled, support-bound, third hybrid complex;

(g) thereafter separating materials not bound to the solid support; and

(h) noting the presence of a detectable signal resulting from interaction between the first and second label probe segments in the support-bound, third hybrid complex.

23. The assay of claim 22, wherein the first label probe segment comprises a first enzyme fragment and the second label probe segment comprises a second enzyme fragment, and wherein the first and second enzyme fragments are complementary enzyme fragments which are inactive unless brought together by the presence of target.

24. The assay of claim 22, wherein the first label probe segment comprises an enzyme and the second label probe

segment comprises a cofactor for the enzyme, and wherein the enzyme and cofactor are inactive unless brought together by the presence of target.

5 25. The assay of claim 22, wherein the first label probe segment comprises an enzyme and the second label probe segment comprises a luminescent or fluorescent activator, wherein the enzyme catalyzes the formation of a product, and the activator enhances the detection of the product.

10

 26. In a solution phase sandwich hybridization assay for detecting a nucleic acid analyte in a sample, comprising: (a) binding the analyte through capture extender molecules directly or indirectly to a solid support-bound capture
15 probes; (b) labelling the analyte; and (c) detecting the presence of analyte-associated label on the support,

 the improvement which comprises incorporating into the assay a competitor oligonucleotide containing a nucleic acid sequence which is capable of hybridizing to the solid
20 support-bound capture probes.

 27. A solution phase sandwich hybridization assay for detecting the presence of a nucleic acid analyte in a sample, comprising:

25 (a) providing a solid support having capture probes thereon, capture extender molecules having a first segment C-1 capable of hybridizing to a nucleic acid sequence in the analyte and a second segment C-2 capable of hybridizing to a nucleic acid sequence in the capture probes, competitor
30 oligonucleotides containing a nucleic acid sequence which is capable of hybridizing to the capture probes, label extender molecules having a first segment L-1 capable of hybridizing to a nucleic acid sequence in the analyte and a second segment L-2 capable of hybridizing to a label probe system, a
35 label probe system comprising a nucleic acid sequence M-1 capable of hybridizing to L-2 and which, directly or indirectly, gives rise to a detectable signal;

- 69 -

(b) incubating the nucleic acid analyte under hybridization conditions with the capture extender molecules, competitor oligonucleotides, label extender molecules and the capture probes on the solid support, simultaneously or
5 sequentially in any order, to form (i) a first hybrid complex of capture extender, nucleic acid analyte and label extender, and (ii) capped capture probes which have reacted with the competitor oligonucleotide;

(c) thereafter optionally separating materials not
10 bound to the solid support;

(d) thereafter contacting the support-bound hybrid complex with the label probe system under hybridizing conditions, to produce a labeled, support-bound hybrid complex; and

(e) thereafter separating materials not bound to the
15 solid support; and

(f) detecting the presence of label in the labeled support-bound hybrid complex.

20 28. The assay of claim 27, wherein the label probe system comprises (i) an amplification multimer containing the nucleic acid sequence M-1 and a plurality of identical oligonucleotide subunits containing nucleic acid sequences M-2 capable of hybridizing to label probes, and (ii) label
25 probes containing a nucleic acid sequence L-3 which is capable of hybridizing to M-2 and which, directly or indirectly, gives rise to a detectable signal, and wherein step (d) of the assay comprises the following steps:

(d₁) contacting the support-bound hybrid complex under
30 hybridization conditions with the amplification multimer, to produce a second support-bound hybrid complex;

(d₂) thereafter optionally separating materials not bound to the solid support; and

(d₃) thereafter contacting the second support-bound
35 hybrid complex with the label probes under hybridization conditions, to produce a labeled support-bound hybrid complex.

- 70 -

29. A solution phase sandwich hybridization assay for detecting the presence of a nucleic acid analyte in a sample, comprising:

- (a) providing a solid support having capture probes
5 thereon, capture extender molecules having a first segment C-1 capable of hybridizing to a nucleic acid sequence in the analyte and a second segment C-2 capable of hybridizing to a nucleic acid sequence in the capture probes, competitor
10 oligonucleotides containing at least one nucleic acid sequence which is capable of hybridizing to the capture probes, label extender molecules having a first segment L-1 capable of hybridizing to a nucleic acid sequence in the analyte and a second segment L-2 capable of hybridizing to a nucleic acid sequence P-1 in a preamplifier probe, a
15 preamplifier probe having the nucleic acid sequence P-1 and capable of binding a plurality of amplification multimers through a nucleic acid sequence P-2, amplification multimers containing a nucleic acid sequence M-1 capable of hybridizing to P-2 and a plurality of identical oligonucleotide subunits
20 containing nucleic acid sequences M-2 capable of hybridizing to label probes, and label probes containing a sequence L-3 which is capable of hybridizing to M-2 and which, directly or indirectly, gives rise to a detectable signal;

- (b) incubating the nucleic acid analyte under
25 hybridization conditions with the capture extender molecules, competitor oligonucleotides, label extender molecules and the capture probes on the solid support, simultaneously or sequentially in any order, to form (i) a first hybrid complex of capture extender, nucleic acid analyte and label extender,
30 and (ii) capped capture probes which have reacted with the competitor oligonucleotide;

(c) thereafter optionally separating materials not bound to the solid support;

- (d) contacting the support-bound, first hybrid complex
35 under hybridization conditions with the preamplifier probe and the amplification multimers, to produce a support-bound, second hybrid complex;

- 71 -

(e) thereafter optionally separating materials not bound to the solid support;

(f) contacting the support-bound, second hybrid complex with the label probes under hybridizing conditions, to
5 produce a labeled, support-bound, third hybrid complex;

(g) thereafter separating materials not bound to the solid support; and

(h) detecting the presence of label in the support-bound, third hybrid complex.

10

30. A kit for detecting a nucleic acid analyte in a sample, comprising:

(a) a solid support having capture probes bound thereto;

15

(b) a first capture extender molecule capable of hybridizing to the capture probes and to predetermined segments of the nucleic acid analyte;

20

(c) a distinct second capture extender molecule capable of hybridizing to the capture probes and to segments of the nucleic acid analyte other than those to which the first capture extender molecule binds;

(d) label extender molecules capable of hybridizing to segments of the nucleic acid analyte other than those to which the first and second capture extender molecules bind;

25

(e) a label probe system containing a nucleic acid sequence capable of hybridizing to the label extender molecules and which provide, directly or indirectly, a detectable signal

wherein the capture probes comprises a probe having a
30 first capture extender binding sequence capable of hybridizing to the first capture extender molecule and a second capture extender binding sequence capable of hybridizing to the second capture extender molecule.

35

31. The kit of claim 30, wherein the label probe system comprises (f) an amplification multimer containing a nucleic acid sequence capable of hybridizing to the label

extender molecules and a plurality of identical oligonucleotide subunits; and

- (g) label probes designed to hybridize to the identical oligonucleotide subunits and which provide, directly or indirectly, a detectable signal.

32. A kit for detecting a nucleic acid analyte in a sample, comprising:

- (a) a solid support having capture probes bound thereto;
- (b) capture extender molecules capable of hybridizing to the capture probes and to predetermined segments of the nucleic acid analyte;
- (c) a first label extender molecule capable of hybridizing to predetermined segments of the nucleic acid analyte;
- (d) a second label extender molecule capable of hybridizing to segments of the nucleic acid analyte other than those to which the first label extender molecule binds;
- (e) an optional preamplifier probe capable of binding to the label extender molecules and to a plurality of amplification multimers;
- (f) an amplification multimer containing a nucleic acid sequence capable of hybridizing to the label extender molecules or to the preamplifier probe, and a plurality of identical oligonucleotide subunits; and
- (g) label probes designed to hybridize to the identical oligonucleotide subunits and which provide, directly or indirectly, a detectable signal.

33. A kit for detecting a nucleic acid analyte in a sample, comprising:

- (a) a solid support having capture probes bound thereto;
- (b) capture extender molecules capable of hybridizing to the capture probes and to predetermined segments of the nucleic acid analyte;

(c) a first label extender molecule capable of hybridizing to predetermined segments of the nucleic acid analyte;

5 (d) a second label extender molecule capable of hybridizing to segments of the nucleic acid analyte other than those to which the first label extender molecule binds;

(e) a first amplification multimer containing (i) a nucleic acid sequence capable of hybridizing to the first label extender molecule, and (ii) a plurality of identical
10 oligonucleotide subunits;

(f) a second amplification multimer containing (i) a nucleic acid sequence capable of hybridizing to the second label extender molecule, and (ii) a plurality of identical oligonucleotide subunits; and

15 (g) label probes containing a first nucleic acid sequence capable of hybridizing with the oligonucleotide subunits of the first amplification multimer, and a second nucleic acid sequence capable of hybridizing with the oligonucleotide subunits of the second amplification
20 multimer, and which provide, directly or indirectly, a detectable signal.

34. A kit for detecting a nucleic acid analyte in a sample, comprising:

25 (a) a solid support having capture probes bound thereto;

(b) capture extender molecules capable of hybridizing to the capture probes and to predetermined segments of the nucleic acid analyte;

30 (c) a competitor oligonucleotide containing a nucleic acid sequence capable of hybridizing to the capture probes;

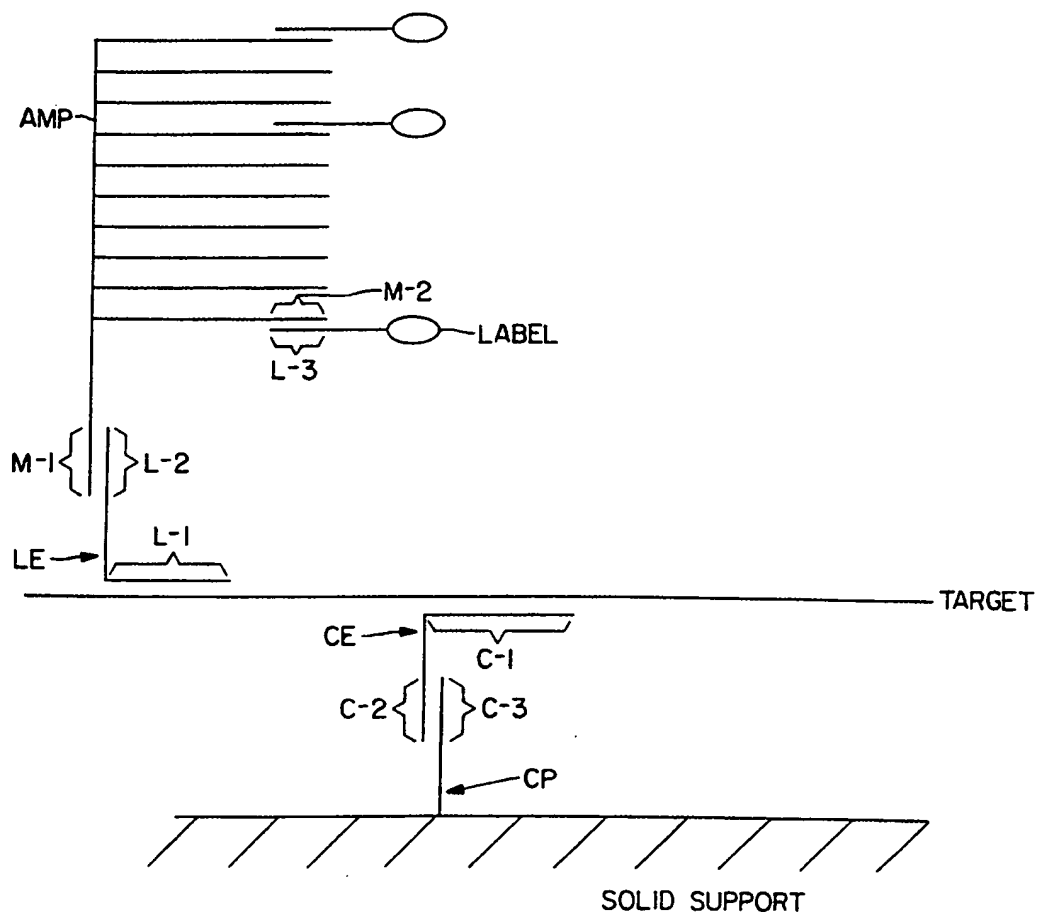
(d) label extender molecules capable of hybridizing to segments of the nucleic acid analyte other than those to which the capture extender molecules bind;

35 (e) an optional preamplifier probe capable of binding to the label extender molecules and to a plurality of amplification multimers;

(f) an amplification multimer containing a nucleic acid sequence capable of hybridizing to the label extender molecules or to the preamplifier probe, and a plurality of identical oligonucleotide subunits; and

- 5 (g) label probes designed to hybridize to the identical oligonucleotide subunits and which provide, directly or indirectly, a detectable signal.

FIG. 1



2 / 9

FIG. 2

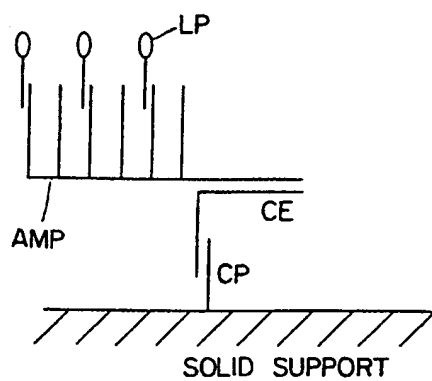


FIG. 3

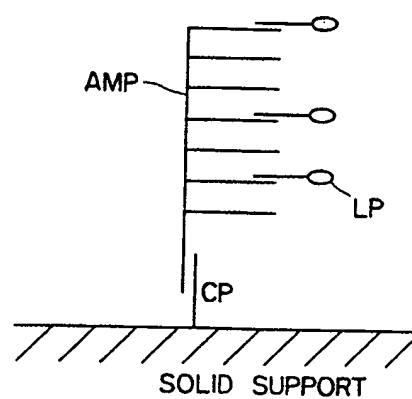


FIG. 4

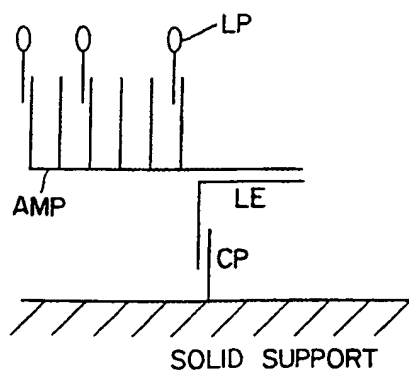
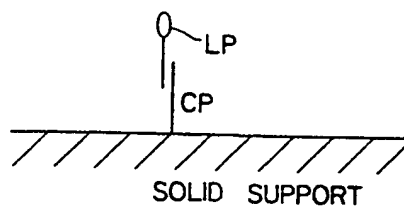


FIG. 5



3/9

FIG. 6

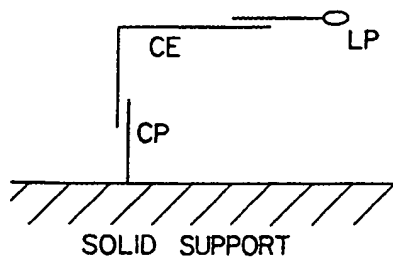


FIG. 7

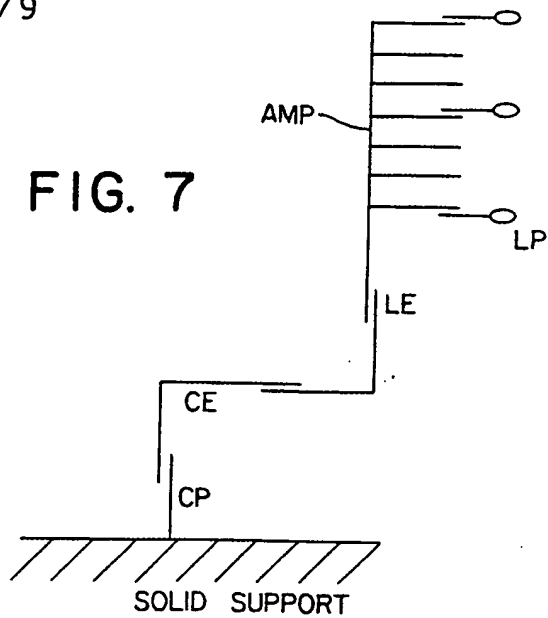
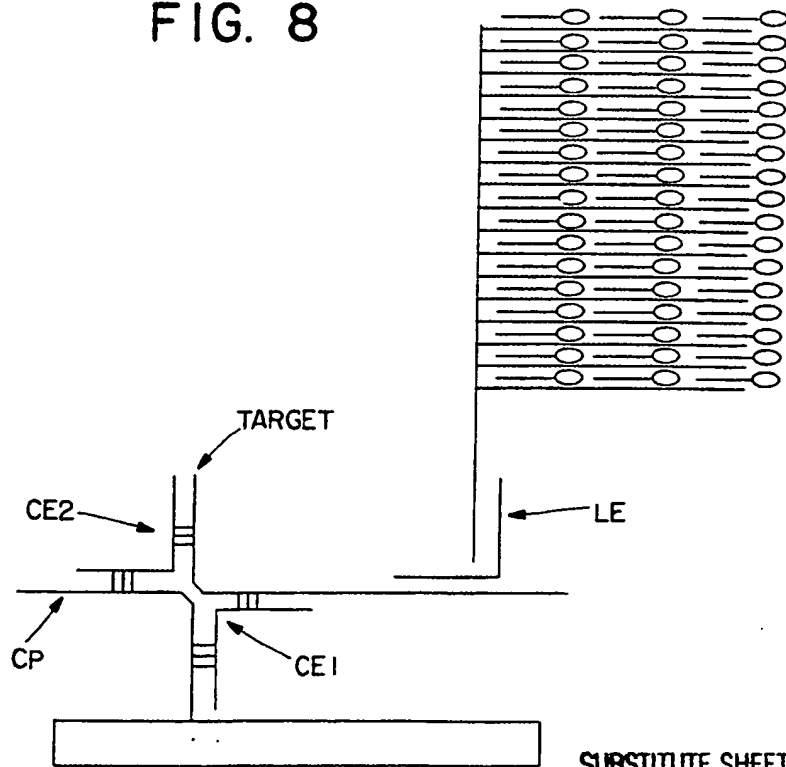


FIG. 8



4 / 9

FIG. 9

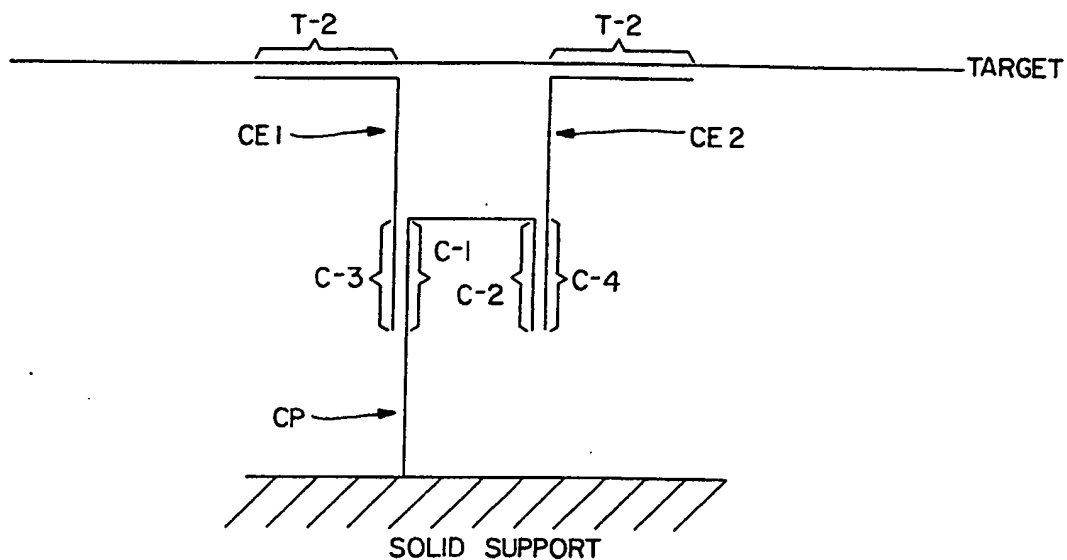
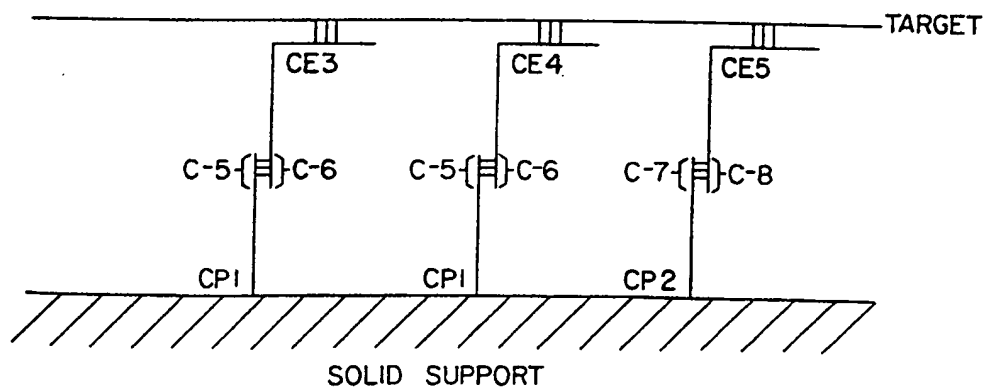
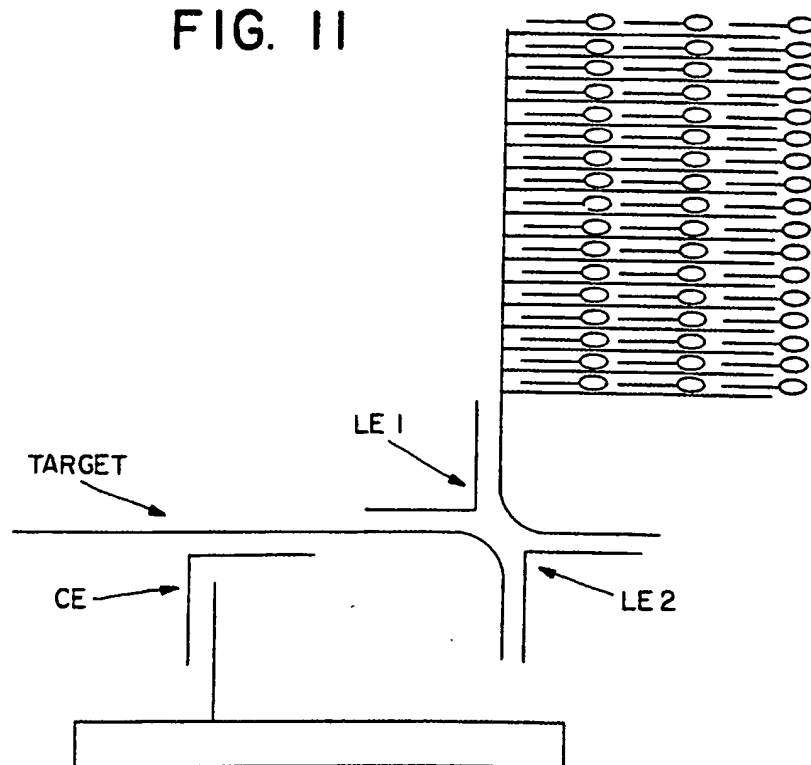


FIG. 10



5/9

FIG. II



6 / 9
FIG. 12

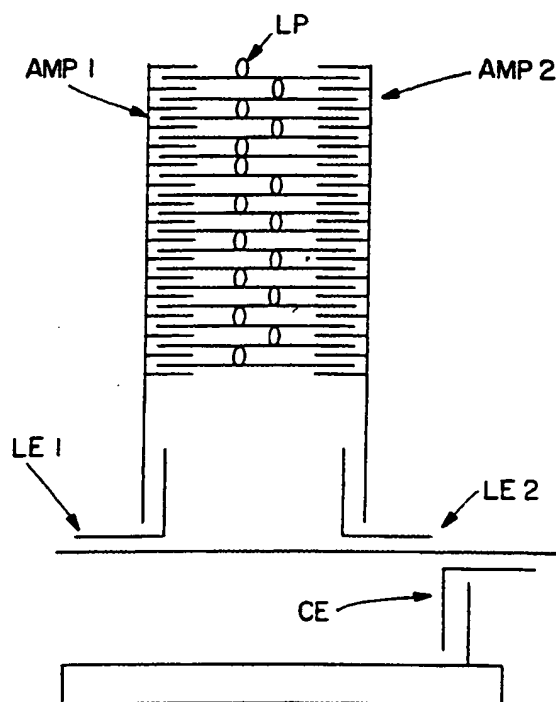
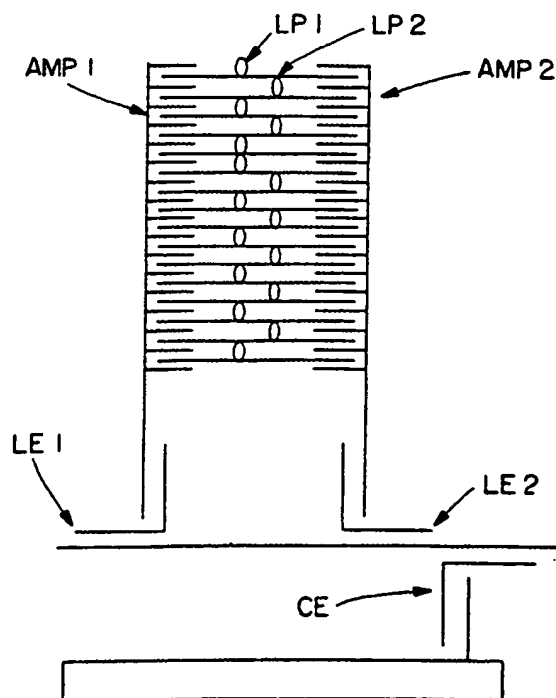


FIG. 13



7/9

FIG. 14

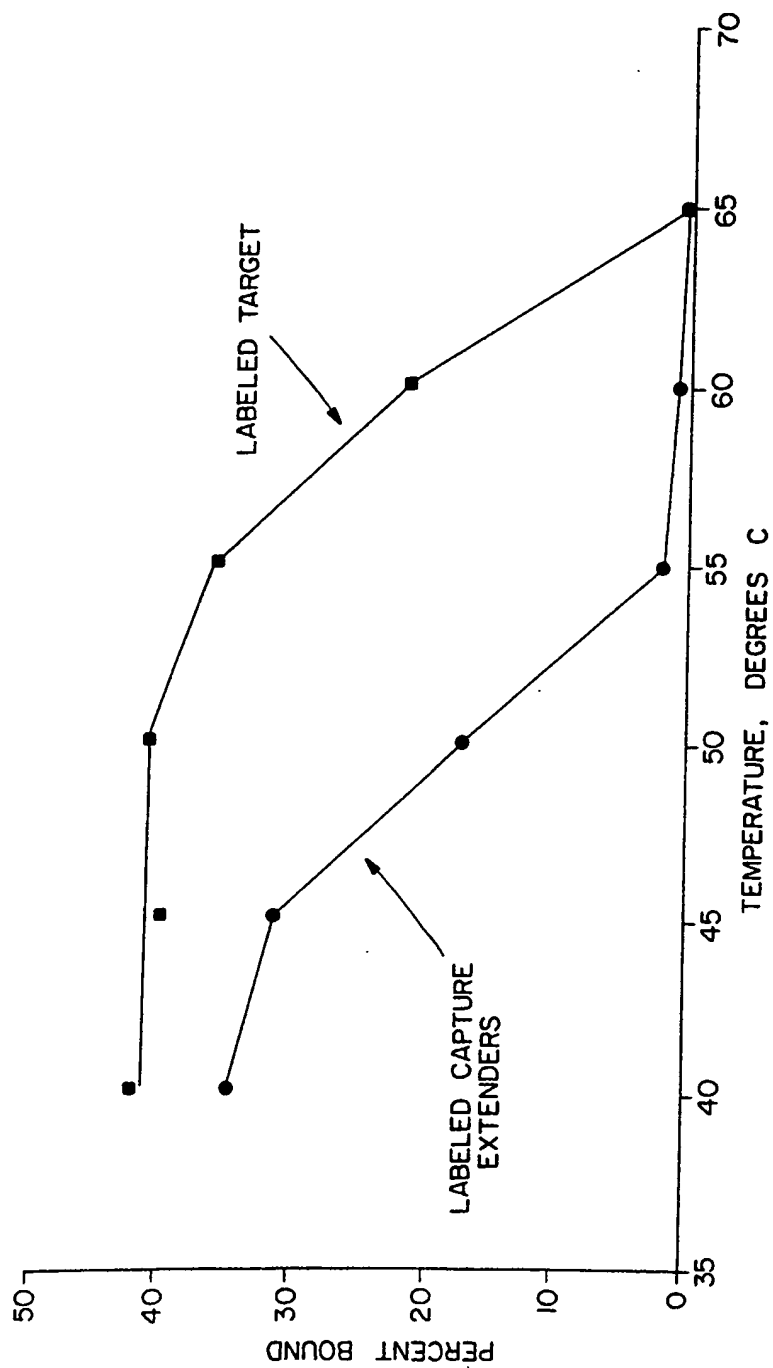
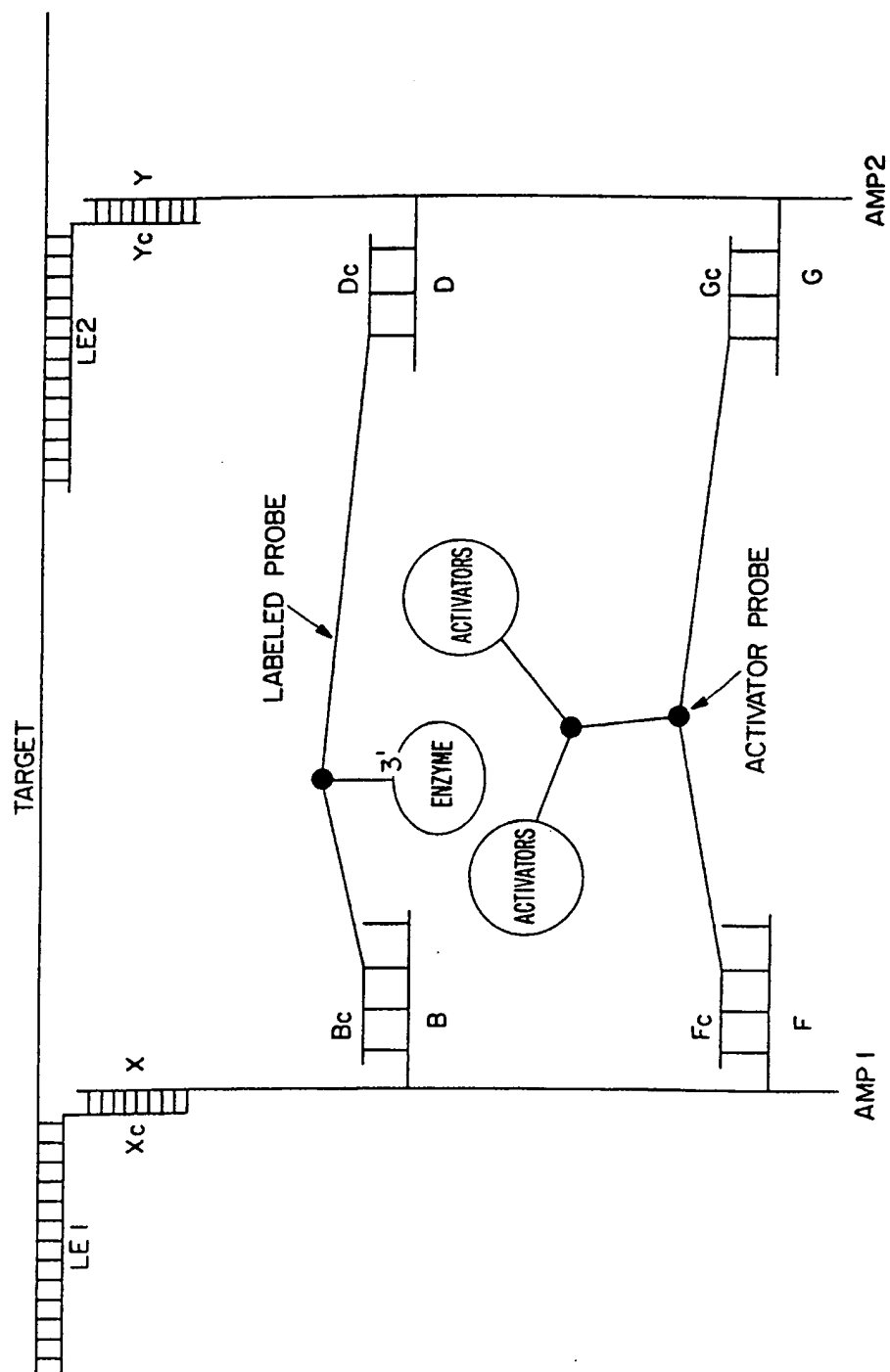


FIG. 15

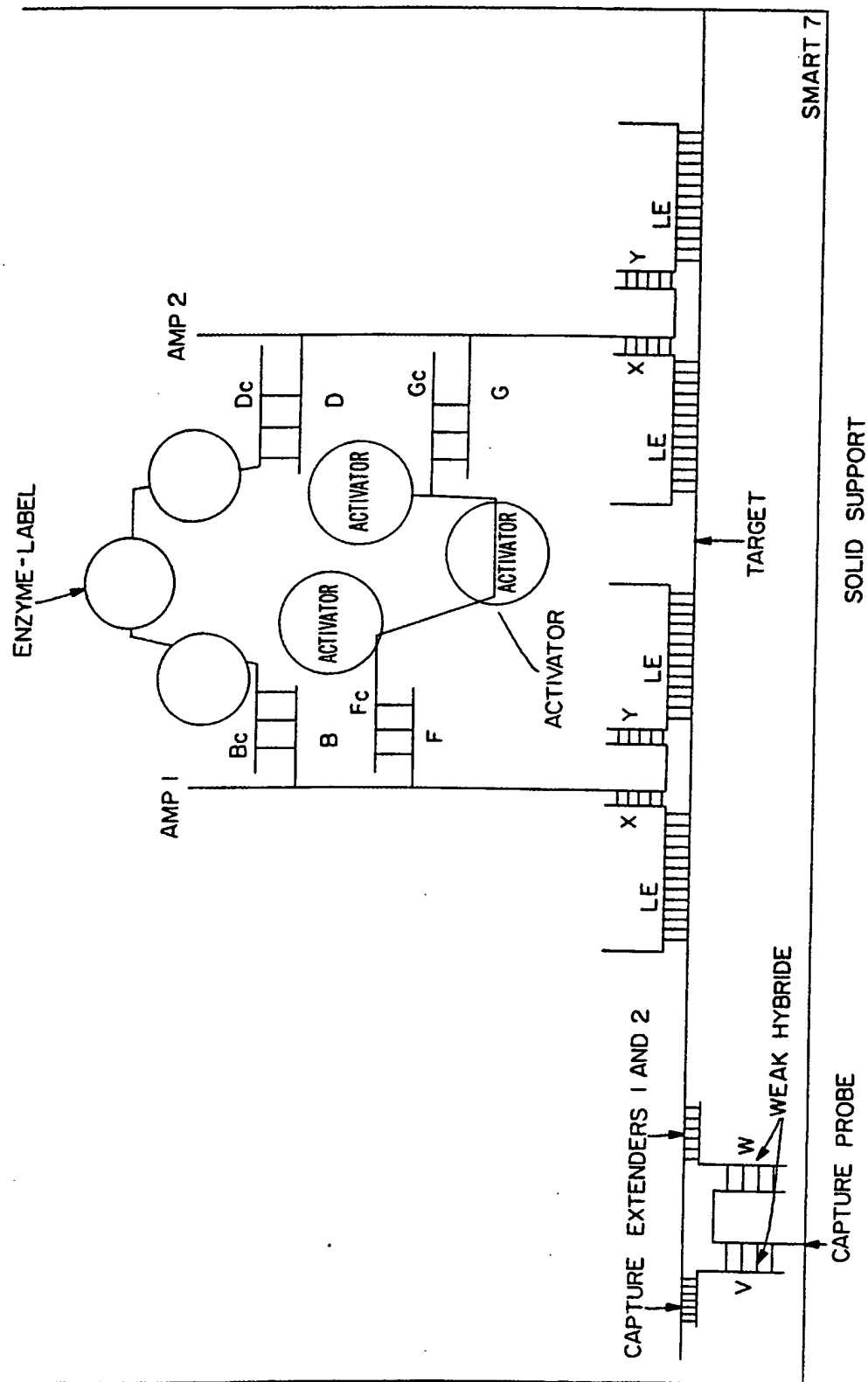
X AND Y FROM STABLE HYBRIDS; B, D, F, G, DO NOT FORM STABLE HYBRIDS



SMART 3a

FIG. 16

B, D, F, G, V, W, X, Y: ALL FORM UNSTABLE HYBRIDES



INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 94/14119

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12Q1/68 C07H21/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C12Q C07H

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO,A,93 13221 (CHIRON CORPORATION) 8 July 1993 see page 5 see page 9, last paragraph - page 10, paragraph 1; claims 16,17 ----	1-34
X	WO,A,93 13223 (CHIRON CORPORATION) 8 July 1993 see the whole document ----	1-34
X	WO,A,93 13227 (CHIRON CORPORATION) 8 July 1993 see the whole document ----	1-34
A	EP,A,0 469 755 (SYNTEX (U.S.A) INC.) 5 February 1992 see page 13 - page 15 -----	

☐ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "B" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- "&" document member of the same patent family

Date of the actual completion of the international search

27 April 1995

Date of mailing of the international search report

11-05-1995

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax (+31-70) 340-3016

Authorized officer

Espen, J

INTERNATIONAL SEARCH REPORT

Information on patent family members

Internat Application No

PCT/US 94/14119

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
WO-A-9313221	08-07-93	AU-B-	3467293	28-07-93
		CA-A-	2124795	08-07-93
		JP-T-	7502414	16-03-95
WO-A-9313223	08-07-93	JP-T-	7502654	23-03-95
WO-A-9313227	08-07-93	EP-A-	0625214	23-11-94
		JP-T-	7502655	23-03-95
		US-A-	5407795	18-04-95
EP-A-0469755	05-02-92	CA-A-	2047342	20-01-92
		JP-A-	5084079	06-04-93

**This Page is Inserted by IFW Indexing and Scanning
Operations and is not part of the Official Record**

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images include but are not limited to the items checked:

- ☐ BLACK BORDERS
- ☐ IMAGE CUT OFF AT TOP, BOTTOM OR SIDES
- ☐ ~~FADED TEXT OR DRAWING~~
- ☐ BLURRED OR ILLEGIBLE TEXT OR DRAWING
- ☐ SKEWED/SLANTED IMAGES
- ☐ COLOR OR BLACK AND WHITE PHOTOGRAPHS
- ☐ GRAY SCALE DOCUMENTS
- ☐ ~~LINES OR MARKS ON ORIGINAL DOCUMENT~~
- ☐ REFERENCE(S) OR EXHIBIT(S) SUBMITTED ARE POOR QUALITY
- ☐ OTHER: _____

IMAGES ARE BEST AVAILABLE COPY.

As rescanning these documents will not correct the image problems checked, please do not report these problems to the IFW Image Problem Mailbox.